



Protective Effect of *Boswellia carteri* on Aluminium Chloride-Induced Alzheimer's Disease in Male Albino Rat

A. E. Bawazir*, R. M. Aljarari

Department of biology, Faculty of Science, Jeddah University, Jeddah, Saudi Arabia.

ABSTRACT

Alzheimer's disease (AD) is one of the most common dementia causes. *Boswellia carteri* is used for its memory enhancing effects. The present experimental study investigates the effects of *B. carteri* on aluminium chloride (AlCl_3)-induced AD in adult male albino rats. Forty adult male Albino rats were randomly divided into five groups (8 rats each). Rats of the first group were served as controls; rats of the second group *B. carteri* group were supplemented orally with *B. carteri* aquatic extract (500 mg /kg b.w./day) for 8 weeks; rats of the third group AlCl_3 group (AD model) were treated orally with AlCl_3 (100 mg/ kg b.w./day) for 8 weeks; rats of the forth group (*B. carteri* + AlCl_3) received 500 mg/ kg b.w./day and AlCl_3 (100 mg/ kg b.w./day) orally for 8 weeks; rats of the fifth group (rivastigmine + AlCl_3) received orally rivastigmine (0.3 mg/kg.w./day) and AlCl_3 (100 mg/kg b.w./day) for 8 weeks. After eight weeks, the behavioral test (maze learning test) was done for rats to assess learning and memory, acetylcholinesterase (AChE), some neurotransmitter levels [dopamine (DA), norepinephrine (NE), gamma amino butyric acid (GABA)] and oxidative stress markers as [reduced glutathione (GSH), oxidase glutathione (GSSG), superoxide dismutase (SOD) and lipid peroxidation (LPO)] were measured in cortex and hippocampus homogenate, and histopathological studies were made for the hippocampus area. Aluminum exposure significantly decreased the learning and memory in the maze learning test. Significant increase of cortex and hippocampus homogenate levels of AChE and LPO but significant decrease in NE, DA and GABA, GSH, GSSG and SOD were observed in rats subjected to AlCl_3 . Histopathological examination of hippocampus sections showed severe changes including increase of degenerated cells and structural damage in AlCl_3 treated rats. Treatment of rats with *B. carteri* or rivastigmine leads to improvement of rat's memory and learning, neurotransmitters and oxidative stress markers and pronounced attenuation on the hippocampus and cortex damage caused by AlCl_3 . This study suggests that chronic oral intake of *B. carteri* have neuroprotective effect and improve the learning and memory in AD rat's models and these effects may be due to its antioxidant properties.

Key Words: Alzheimer's Disease, Aluminium Chloride, *Boswellia Carteri*, Maze Learning Test, Acetylcholinesterase, Neurotransmitters, Oxidative Stress Markers, Rats.

eIJPPR 2018; 8(6):29-39

HOW TO CITE THIS ARTICLE: E. Bawazir, R. M. Aljarari (2018). "Protective Effect of *Boswelliacarteri* on Aluminium Chloride-Induced Alzheimer's Disease in Male Albino Rat", International Journal of Pharmaceutical and Phytopharmacological Research, 8(6), pp.29-39.

INTRODUCTION

The world's population is rapidly aging which is considered as a multi-factorial process and causes several changes from cellular to organism levels. Nowadays, the primary focus has been on the use of higher cognitive ability such as working memory, long-term memory, mental images or reinforcing learning, judgment and reasoning in cognitive neuroscience [1]. Alzheimer's disease (AD) is neurodegenerative disease that results in

complete need for care after diagnosis. It is a main reason of dementia in people aged above 60 years [2]. Alzheimer's disease (AD) is the most common form of chronic diseases among the elderly [3]. It can be difficult to differentiate between the different types of dementia using laboratory tests only, as not all of the causes of dementia can be identified [4]. AD has become a major medical, social and financial burden. AD results from multifactorial processes including environmental exposure, genetic risk factors, cerebrovascular risk

Corresponding author: A. E. Bawazir

Address: Department of Biology, Faculty of Science, Jeddah University, Jeddah, Saudi Arabia .

E-mail: aeabawazir@uj.edu.sa

Relevant conflicts of interest/financial disclosures: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest .

Received: 02 September 2018; **Revised:** 23 November 2018; **Accepted:** 26 November 2018



factors, oxidative stress, infectious agents, age and sex [5]. Many of neurotransmitters are disturbed in AD as the cholinergic, adrenergic, glutamatergic, serotonergic, GABAergic and dopaminergic neurotransmitters [6].

The human organisms are constantly and inevitably exposed to aluminium (Al). Al increases as a result of continued acidification of industrialized society environment and it causes harmful effects on various organs. Al is the highly neurotoxic element and its exposure is proposed to be included in AD development where it produce clinical and pathological features similar to AD. There is no curative treatment for AD and current pharmacological treatments only gives temporarily symptomatic relief in some patients [7].

Many studies have shown that herbal medicine has the ability to provide effective treatment of neurodegenerative diseases as AD where natural compounds have antioxidant properties and very low side effects. For about 3000 years, *Boswellia* tree (frankincense) resin was an important trade material in North Africa and Arabian Peninsula. In Arabic language, frankincense is also known as “al-luban”, which means “white” or “cream” and is a basis of the other name, olibanum [8]. Frankincense contains 6-30% gums (polysaccharides), 60-85% resins (terpenes), and 5-9% essential oil [9]. It had been reported that frankincense from many species of *Boswellia* genus improves memory in both normal brain [10, 11] and affected memory cases [12, 13].

Relatively few studies have examined the effect of *B. carteri* on AD. This study aimed to examine the protective actions of chronic oral intake of *B. carteri* on AD rat's model.

MATERIALS AND METHODS

Animals

Forty adult male Albino Wistar rats weighting 170-210 g were used in the study. The rats were obtained from the Experimental Animal Unit of Faculty of Pharmacy, King Abdulaziz University, Jeddah, Saudi Arabia. The animals were housed 8 rats per cage and maintained under controlled laboratory status of 12 h light: dark cycle, 65% humidity and temperature of 20 ± 1 °C. Rats were acclimatized to laboratory status for one week before beginning of experiments. Rats fed *ad libitum* on commercial chow and freely access tap water. All experimental steps were approved and made according to the ethical roles of Animal Care and Use Committee of King Abdulaziz University (1161439).

Chemicals

AlCl_3 (Techno Pharmchem, Haryana, India) was supplied as white powder. Rivastigmine, Exelon, 1.5 mg was

purchased from Novartis Company (Basel, Switzerland). Sodium chloride (NaCl) and sodium hydroxide (NaOH) were obtained from Panreac (Barcelona, Spain). Potassium dihydrogen phosphate (KH_2PO_4), potassium chloride (KCl), disodium phosphate (Na_2HPO_4), and formaldehyde were obtained from Riedel-dehaen (Sleeze, Germany).

Olibanum (*B. carteri*)

B. carteri was obtained from a local market in Jeddah, Saudi Arabia. *B. carteri* was imported from Somalia. It was purchased in closed packages.

Preparation of Olibanum Extraction

A volume of 50 ml of boiling-hot distilled water was poured on 5 g of the resin in a beaker. The mixture was allowed to stand for 30 min before it was filtered with a filter paper. The infusion was always freshly prepared so as to prevent growth of fungi [14].

Experimental Design

The experimental rats were sorted into five experimental groups, 8 rats each. Groups were: **Group 1 (control group):** Rats were orally supplemented with 0.9 ml of saline solution (0.9% NaCl, vehicle) daily for eight weeks. **Group 2 (*B. carteri* group):** rats were orally supplemented with *B. carteri* aquatic extract at a dose 0.9 ml that contains 500 mg/ kg b.w./day for eight weeks [15]. **Group 3 (AlCl_3 group):** rats were given 0.9 ml that contains 100 mg/ kg b.w./day of AlCl_3 orally daily for eight weeks [16]. **Group 4 (*B. carteri* + AlCl_3 group):** rats were orally supplemented with *B. carteri* aquatic extract at the same dose taken to group 2 and AlCl_3 after 1 hour at the same dose given to group 3 daily for eight weeks. **Group 5 (Rivastigmine+ AlCl_3 group):** rats were orally supplemented with rivastigmine at the dose 0.9 ml that contains 0.3 mg/ kg b.w./ day [17] and AlCl_3 after 1 hour at the same dose given to group 3 daily for eight weeks.

Behavior Study (Maze learning test)

Maze learning test is a reliable method of studying trial and error learning. The base measure of the maze was 100 x 60 cm (L x W), and the walls were 20 cm high. The entire maze was made of wood with a glass cover over the maze to prevent escape of rats and to allow easy observation. Rats were deprived of food 23 hours before the initiation of the experiments. Rats were given their daily amount of food as a reward at the end of the maze. The hungry rats were given one trial per day for five consecutive days. The following measurements were recorded: the elapsed time to reach the food (minutes) and the number of errors (passage in blind alleys) as previously described [18].

Tissue preparation

After the behavioral test, the rats were euthanized by decapitation according to the rules of King Abdulaziz University. After dissection, their brains were removed and washed with saline and divided into two halves. One fixed in 10% formalin for histopathological studies. The hippocampus and cortex of the other half was dissected out. Then, hippocampus and cortex tissues were immediately frozen in -80°C for preparation of tissues homogenate. For biochemical studies the frozen hippocampus and cortex were homogenized separately, and centrifuged for 15 min at 3000 revolutions per minute (rpm) at 4°C . After that, supernatant was separated, and aliquot was stored in Eppendorf tube at -80°C for the biochemical assay.

Biochemical assay

ELISA kits for the measurement of hippocampus and cortex tissues homogenate for Acetylcholinesterase (AChE) (Cat. # MBS038896), Norepinephrine (NE) (Catalog # MBS269993), Dopamine (DA) (Catalog Number # MBS7214676) and Gamma-aminobutyric acid (GABA) (Catalog Number # MBS740443), Superoxide Dismutase (SOD) (Catalog # MBS266897), Lipid Peroxide (LPO) (Catalog # MBS2515688) while, Fluorometric Assay kits were used to measure reduced glutathione (GSH) and oxidase glutathione (GSSG) (Catalog Number # MBS841503). All the kits were purchased from My BioSource (San Diego, USA).

Histopathological studies

After fixation of brain tissues in formalin saline (10%) for 24 hours, brain tissues were subjected to serial dilution of alcohol for dehydration. Brain tissues were immersed in paraffin, cut into $3\ \mu\text{m}$ thick sections and stained in haematoxylin and eosine (H and E) to be examined by light microscope.

Statistical analysis

The statistical analysis was made by Statistical Package for Social Science (SPSS program, version 25) (SPSS Inc., Chicago, IL, USA). Values were expressed as mean \pm standard error. The difference between different experimental groups was made using One Way ANOVA (Tukey test). P -value <0.05 was recognized as significant value.

RESULTS

Behavior study (Maze learning test)

At the 1st day, duration of elapsed time in the maze was significantly increased in AlCl_3 group versus control ($P < 0.05$). Compared with AlCl_3 group, duration of elapsed time in the maze was significantly decreased in rivastigmine + AlCl_3 group ($P < 0.05$), while in *B. carteri* + AlCl_3 group was significantly increased. At the 2nd, 3rd, 4th and 5th days, duration of elapsed time in the maze was significantly prolonged in AlCl_3 group compared with control, *B. carteri* + AlCl_3 , and rivastigmine + AlCl_3 groups ($P < 0.05$) (Figure 1).

At the 1st, 2nd, 3rd, 4th, and 5th days, number of error in the maze was significantly decreased in control, *B. carteri* +

AlCl_3 and rivastigmine + AlCl_3 groups versus AlCl_3 group ($P < 0.05$) (Figure 2).

Acetylcholinesterase and Some Neurotransmitter levels in cortex and hippocampus homogenate

Levels of AChE in the cortex and hippocampus homogenate was significantly increased in AlCl_3 group versus control group ($P < 0.05$). Cortex and hippocampus AChE levels were significantly decreased in *B. carteri* + AlCl_3 and rivastigmine + AlCl_3 groups versus AlCl_3 treated group ($P < 0.05$). Compared with control, the levels of DA, NE and GABA in the cortex and hippocampus homogenate was markedly decreased in AlCl_3 treated group ($P < 0.05$). Cortex and hippocampus DA, NE and GABA levels in rats treated with AlCl_3 were significantly decreased compared with *B. carteri* + AlCl_3 and rivastigmine + AlCl_3 groups ($P < 0.05$) (Table 1).

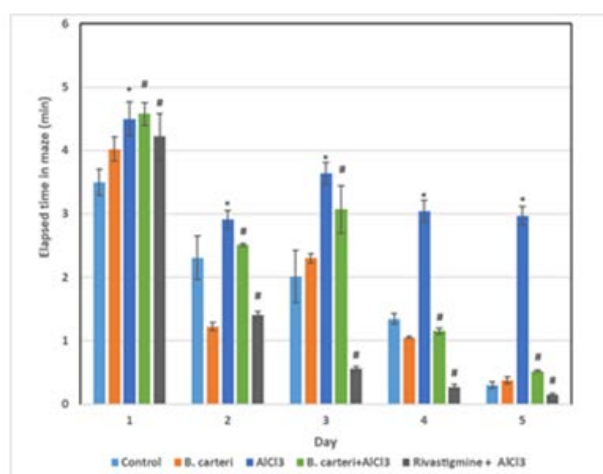


Fig. 1. Protective effects of *Boswellia carteri* and rivastigmine on the time taken to find the food in the learning maze by Alzheimer's disease-induced in rats.

Values were expressed as mean \pm SE

* $P < 0.05$ compared with control # $P < 0.05$ compared with AlCl_3

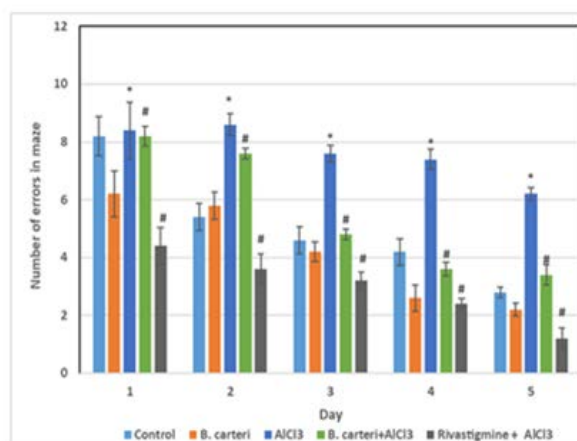


Fig. 2. Protective effects of *Boswellia carteri* and rivastigmine on the number of errors in the learning maze by Alzheimer's disease-induced in rats.

Values were expressed as mean \pm SE.

* $P < 0.05$ compared with control # $P < 0.05$ compared with AlCl_3

Table 1. Protective effects of *Boswellia carteri* and rivastigmine on cortex and hippocampus homogenate levels of acetylcholinesterase and some neurotransmitters content in Alzheimer's disease-induced rats

| Groups Parameters | | Control | <i>B. Carteri</i> | AlCl ₃ | <i>B. Carteri</i> + AlCl ₃ | Rivastigmine + AlCl ₃ |
|-----------------------|--------|---------------|-------------------|----------------------------|---------------------------------------|----------------------------------|
| AChE (U/mg) | Cortex | 0.65±0.0821 | 0.675±0.0567 | 1.3567±0.0761 ^a | 0.6317±0.0956 ^b | 0.7283±0.0624 ^b |
| | Hip | 1.79±0.1099 | 1.675±0.0567 | 2.6883±0.1272 ^a | 1.915±0.0591 ^b | 2.025±0.0615 ^b |
| DA (pg/mg) | Cortex | 3.625±0.2869 | 3.5783±0.1827 | 0.4817±0.1311 ^a | 2.845±0.181 ^b | 3.455±0.2067 ^b |
| | Hip | 4.2±0.4227 | 4.7833±0.3724 | 0.8083±0.1087 ^a | 3.1467±0.1925 ^b | 4.0667±0.249 ^b |
| NE (pg/mg) | Cortex | 4.5867±0.3553 | 4.5833±0.338 | 1.0267±0.3371 ^a | 3.425±0.1954 ^b | 5.8333±0.3566 ^b |
| | Hip | 5±0.4017 | 4.6167±0.3364 | 1.7483±0.2893 ^a | 4.4017±0.2259 ^b | 6.7033±0.2939 ^b |
| GABA (μmol/mg) | Cortex | 0.2767±0.0113 | 0.2918±0.0279 | 0.0738±0.0144 ^a | 0.3052±0.021 ^b | 0.465±0.0184 ^b |
| | Hip | 0.4383±0.0402 | 0.4433±0.035 | 0.0985±0.0142 ^a | 0.38±0.0257 ^b | 0.7067±0.0226 ^b |

Values were expressed as means ±S.E of eight animals. P<0.05 (significant)

a = Significant difference comparing to the normal control group.

b = Significant difference comparing to AlCl₃ group.

Oxidative stress markers levels in cortex and hippocampus homogenate

The levels of GSH, GSSG and SOD were significantly decreased, while LPO was significantly increased in the cortex and hippocampus homogenate of AlCl₃ group versus control (P<0.05). AlCl₃ treated rats with *B. carteri*, and rivastigmine led to significant increase in cortex and hippocampus GSH, GSSG, SOD levels and significant decrease in LPO in *B. carteri* + AlCl₃ and rivastigmine +AlCl₃ groups compared with AlCl₃ group (P<0.05) (Table 2).

Histopathological results

The hippocampus sections in Cornu Ammonis (CA1) were formed of three layers: polymorphic (POL), pyramidal (PYR), and molecular (MOL). Also, the dentate gyrus (DG) was formed of three layers: molecular (MOL), granular (GRA), and polymorphic (POL). The

hippocampus sections in CA1 of control, *B. carteri* extract-treated rats showed normal pyramidal cells with rounded vesicular nuclei. Furthermore, the hippocampus sections in DG of control, *B. carteri* extract treated rats revealed normal granular cells with vesicular nuclei and increase in the thickness of the granular layer in *B. carteri* extract treated group. After eight weeks of AlCl₃ treatment, the treated rats revealed an abnormal morphology in CA1 and DG of hippocampus characterized by degenerated shrunken cells with darkly stained nuclei. In rats treated with *B. carteri* + AlCl₃, rivastigmine + AlCl₃, hippocampus sections in CA1 and DG showed reduced degenerated cells. In addition, the hippocampus cells in CA1 and DG showed minimal changes compared with cells structure of AlCl₃ only treated rats (Figure 3).

32

Table 2. Protective effects of *Boswelliacarteri* and rivastigmine on cortex and hippocampus homogenate levels of oxidative parameters in Alzheimer's disease-induced rats

| Groups Parameters | | Control | <i>B. Carteri</i> | AlCl ₃ | <i>B. Carteri</i> + AlCl ₃ | Rivastigmine + AlCl ₃ |
|-----------------------|--------|----------------|-------------------|-----------------------------|---------------------------------------|----------------------------------|
| GSH (μmol/mg) | Cortex | 4.255±0.3425 | 4.59±0.1627 | 0.8783±0.1126 ^a | 2.9067±0.2153 ^b | 4.2833±0.3722 ^b |
| | Hip | 4.42±0.4694 | 4.1167±0.3331 | 0.7167±0.1126 ^a | 3.0033±0.2966 ^b | 5.725±0.2722 ^b |
| GSSG (μmol/mg) | Cortex | 4.635±0.3059 | 4.905±0.3641 | 1.0417±0.1551 ^a | 3.0083±0.1287 ^{a,b} | 4.9717±0.404 ^b |
| | Hip | 4.99±0.2762 | 4.555±0.3133 | 1.2867±0.1991 ^a | 3.0833±0.0995 ^{a,b} | 5.455±0.2796 ^b |
| SOD (μmol/mg) | Cortex | 85.8333±7.0067 | 91.8333±6.3293 | 20.45±2.7835 ^a | 55.0833±3.7426 ^{a,b} | 91±5.2409 ^b |
| | Hip | 86±6.4057 | 92.4167±7.9185 | 16.2333±2.3638 ^a | 49.9±1.6488 ^{a,b} | 79±6.2794 ^b |
| LPO (μmol/mg) | Cortex | 0.735±0.0703 | 0.9383±0.0961 | 9.2±0.0394 ^a | 2.7467±0.0365 ^b | 0.2483±0.0904 ^b |
| | Hip | 1.0917±0.0703 | 3.3167±0.0961 | 36±0.0394 ^a | 9.645±0.0365 ^{a,b} | 0.7383±0.0904 ^b |

Values were expressed as means ±S.E of eight animals. P<0.05 (significant)

a = Significant difference comparing to the normal control group.

b = Significant difference comparing to AlCl₃ group

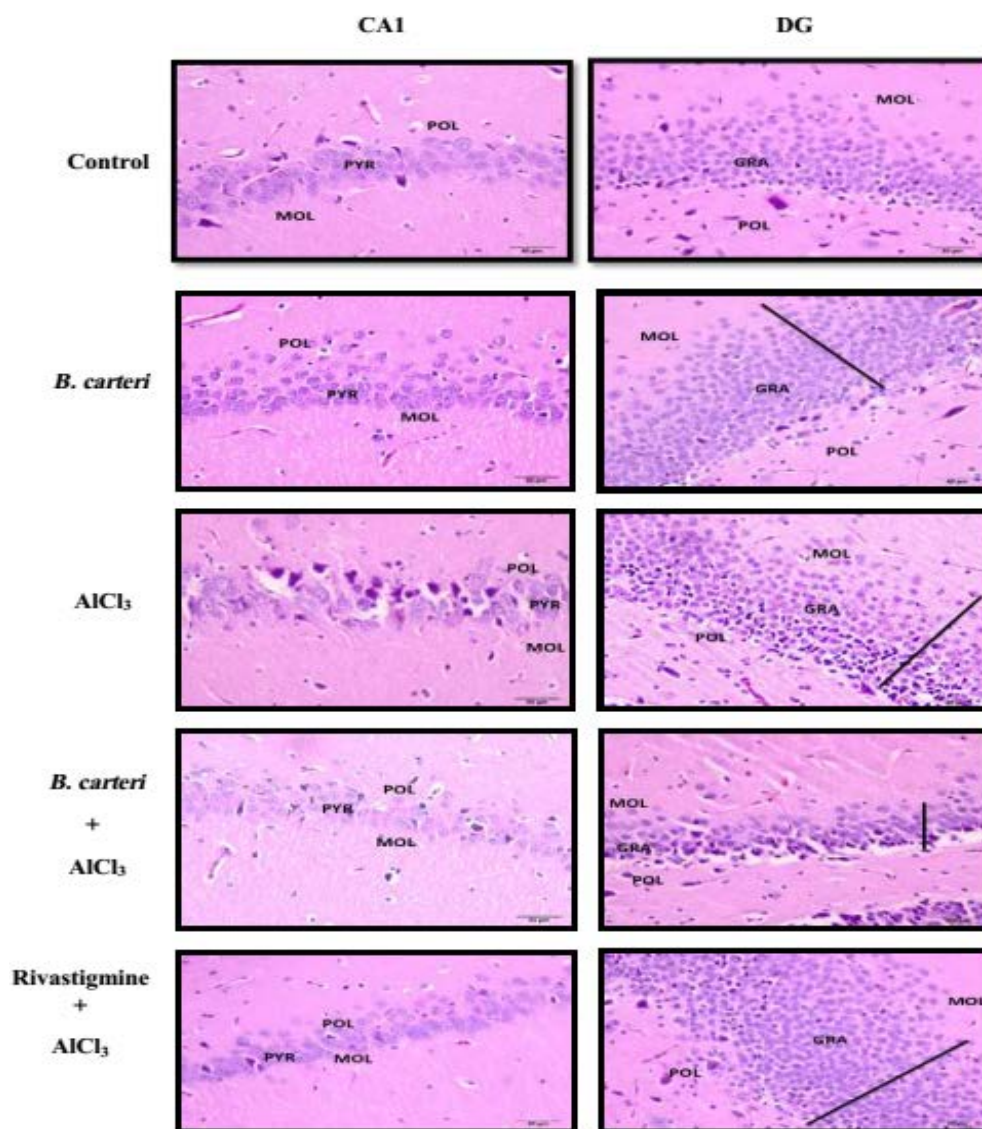


Fig. 3. H&E staining (magnification 400×). The pyramidal cells in CA1 have large vesicular nuclei in control, *B. carteri*-treated groups and rivastigmine treated group but in Al group have irregular forms. The granular cells in DG have vesicular nuclei in control, *B. carteri*-treated groups and rivastigmine treated group but in Al group have irregular forms. Also, an increase was observed in the thickness of the granular layer in *B. carteri*-treated groups and rivastigmine treated group. Treatment by *B. carteri* improved morphological changes induced by $AlCl_3$

DISCUSSION

The results of this study revealed that intake of $AlCl_3$ (100 mg/ kg b.w. orally daily) for eight weeks to rats (AD model) leads to impairment of the learning and memory abilities compared with normal rats that manifested by increase in elapse time and increase in the number of error in the maze learning test to get the food. These results indicated that $AlCl_3$ exposure resulted in impairment of locomotor activity and exploratory behavior (maze learning test) of the rats and decrease in spatial memory. These results agreed with the previous study which reported that Al exposure had a

neurodegeneration effect that leads to learning deficits. Rats injected intraperitoneal with $AlCl_3$ for 60 days (100 mg/ kg b.w.) showed a decrease in the memory in the Morris water task (MWT) and passive avoidance test that assess spatial memory [19]. Ach is responsible to short-term memory. In Al exposed animals, Al can induce disorder in cholinergic neurotransmission leading to memory alterations [20]. Results of this study revealed that, compared with $AlCl_3$ -treated rats, the learning and memory abilities were significantly improved in AD rats group treated with *B. carteri* (500 mg/ kg b.w., orally) daily for 8 weeks as showed by decrease in elapse time, and decrease in the number of errors in the maze.

Therapeutic effect of *B. carteri* on AD rat model caused by streptozotocin (STZ) injection intracerebroventricular (i.c.v) has been studied previously [21] which clarified that STZ increased stimulations numbers needed for initiation of short-term memory and decreased step-through latency (STL) on tested day. Furthermore, oral administration of *B. carteri* aqueous extract (50 mg/kg) for 21 days didn't change learning measurements, but its injection for 42 days significantly enhanced STL, decreased STL number into dark compartment and decreased stay time in dark compartment. Also, Zaker *et al.* [22] found that acute administration of the aqueous extract of *B. carteri* (100 and 300 mg/kg b.w.) orally 30 minutes before the test didn't change learning, but chronic administration of 50 mg/kg b.w. for 21 days improved memory retrieval. Beheshti and Karimi [23] studied the effect of *B. carteri* on memory in rats treated with lipopolysaccharide (LPS) that disturbed memory retrieval by significantly decreasing STL and increasing tumor necrosis factor alpha (TNF- α) levels in the hippocampus versus control. Intake of hydro-alcoholic extracts of *B. carteri* (50 mg/kg b.w.) orally prior to LPS improved memory retrieval and reduced hippocampal TNF- α level than control group. Yassin *et al.* [14] revealed that intake of aqueous infusions of *B. serrata* (90 mg/kg/ b.w. daily for 2 weeks) to rats caused a reduction in the duration taken by rats to reach food in T-maze test; in other words, it corrected the cognitive functions and memory in rats. Hosseini *et al.* [12] showed that chronic intake of 100 and 500 mg/ kg *B. serrata* in hypothyroid male rats inhibits deterioration of learning and memory. The action of frankincense on memory is due to structural changes in brain neuronal circuits [24].

Rivastigmine is used as standardized drug for treatment of AD. The rivastigmine efficacy in dementia treatment has been examined in moderate-to-severe AD patients lived in long-term care facilities [25]. Treatment of AD rats with rivastigmine as a protective or therapeutic agent led to significant decrease in error numbers in the maze versus AD-induced groups of rats [14].

It is reported that Ach has essential activity in learning and memory neurons. Cognitive dysfunction was closely related to the decrease of the activity of cholinergic transmitters. AchE is a hydrolase enzyme that plays a key role in cholinergic transmission by catalyzing Ach hydrolysis [26]. The results of the present work showed that oral intake of AlCl₃ for 8 weeks in AD rat model produced a significant increase in AchE activities of the cerebral cortex and hippocampus homogenate accompanied with a significant decrease in DA, NE and GABA contents in comparison with control group. AchE enzyme has received marked attention in Al neurotoxicity studies [27]. Such results of increase of AchE activities

were coincided with the findings of others [14, 28]. Also, Kaizer *et al.* [29] and Zhang *et al.* [30] demonstrated an increase in AchE activity in different mice and rats' brain regions after treatment with Al. It has been demonstrated that the increase in AchE activity after Al exposure was due to allosteric action between Al and peripheral anionic location of enzyme molecule resulting in modification of secondary structure and enzyme activity [31].

Imbalance in brain bipterins may lead to a disturbance of various neurotransmitters as DA and NE [32]. However, the decrease of monoamines level may be due to alterations in the dopaminergic system [33], or might be due to decreased synthesis resulting from Al damage of the ileal mucosa and reduction in ilea absorption, where a decrease in tryptophan absorption would decrease serotonin synthesis, and a decrease in L-tyrosine absorption may decrease DA, NE and epinephrine (E) production [34]. Moreover, the marked changes of brain neurotransmitters levels (DA and NE) in AlCl₃ rats may be due to enhanced formation of O₂⁻ and H₂O₂ in the hippocampus that leads to neurodegenerative diseases and Lewy bodies' accumulation [35]. Furthermore, the increased iron species concentration among Al rats groups enhance DA oxidation forming DA quinones that react with cysteine residues of functional proteins as GSH suppressing their activities and detoxification actions [36].

Regarding inhibitory neurotransmitters, AlCl₃ induced reduction in cerebral cortex and hippocampus GABA contents, observed herein, is similar to that described previously [37] who demonstrated that AlCl₃ selectively modulates GABA receptors function leading to widespread changes in inhibitory circuits that contribute to neuropathology [38]. The significant decline in GABA level may be due to its increased destruction via enzymatic action and selective loss of GABAergic neurons due to exposure to AlCl₃ [39]. Generally, disturbance in amino acids neurotransmitters leads to direct or indirect AlCl₃ effects on protein metabolism and interfere with GABA formation and destruction via a reactions known as GABA shunt. Both GABA and glutamate may have some common ways of metabolism including astrocyte where they are up taken and changed to glutamine, which is then returned back to the presynaptic vesicles. In the excitatory neurons it is changed to glutamate while in the inhibitory neurons it is changed to GABA with glutamate as intermediate substance [28]. Meanwhile, Shuchanga *et al.* [40] reported that Al-exposed rats had a lower glutamate levels and higher GABA level.

The results of this study showed that oral intake of *B. carteri* (500 mg/kg b.w., orally) every day for 8 weeks leads to a marked decrease in AchE contents of

hippocampus and cerebral cortex homogenate accompanied with a significant increase in DA, NE and GABA contents in comparison with control group. Yassin *et al.* [14] reported that *B. serrata* infusion extract leads to significant improvement in animals with AD and increase the brain Ach levels and also significantly decrease brain AchE levels in dose dependent manner. Boswellic acids, as the major components of *B. serrata* are reported to have anticholinesterase activity [41]. It has been demonstrated that 11-keto-beta-boswellic acid and 11alpha-hydroxy-beta-boswellic acid extracted from *B. serrata* exert inhibitory activity on AchE. The suppressive effect of these compounds on AchE appears to be associated with the presence of either free hydroxyl group or keto group at C-11 and the presence of free hydroxyl group at C-3 in the ursane skeleton [42]. Inhibiting neuronal AchE activity would increase Ach level and improve cognitive function [43].

In view of our results, it has been demonstrated that the treatment of AD- rats with rivastigmine produces marked decline in AchE activity of cortex and hippocampus homogenate and increase in DA, NE and GABA levels. These results are in agreement with others who demonstrated that rivastigmine intake increases levels of Ach and inhibits AchE activity in the brain [14]. Rivastigmine is a carbamate derivative pseudo-irreversible cholinesterase inhibitor that suppressed both BuchE1 and AchE. Rivastigmine is destructed by synapse rather than by hepatic cytochrome enzymes [44]. The glutamatergic system is changed after AchE inhibition after rivastigmine [45]. Therefore, the mechanism by which rivastigmine could improve the cognition of these rats is related to its potential to increase Ach and decrease AchE activity as shown in the present study.

In this study, exposure of rats to $AlCl_3$ leads to significant decrease in the cortex and hippocampus homogenates of antioxidant enzymes as GSH, GSSG and SOD and significant increase in oxidative stress as LPO levels. Several studies revealed that GSH decrease is important in oxidative stress production. Decrease of GSH enhances signaling processes, leading to central excitabilities and neuronal death [46]. Deloncle *et al.* [47] reported that $AlCl_3$ neurotoxicity due to increase in LPO and blood-brain barrier damage. SOD catalyzes the rapid removal of superoxide radicals generating hydrogen peroxide (H_2O_2), which is eliminated by catalase (CAT) [48]. The observed decrease in antioxidant enzymes activity after $AlCl_3$ administration versus control groups is in accordance with those reported by others [49]. Mahdy *et al.* [50] reported that $AlCl_3$ AD-induced rats showed significant increases in serum levels of NO and MDA and marked decreases in total antioxidant capacity (TAC) and SOD activities. Abd-Elhady *et al.* [51] reported that after Al

toxicity, GSH synthesis was declined due to glutathione synthetase decrease. Oxidative stress with the subsequent production of reactive oxygen species (ROS) was postulated as one of the mechanisms of Al toxicity [52]. Free radicals cause protein and DNA injury, inflammation that lead to cellular apoptosis. Moreover, Al disturbs the antioxidant mechanism of eliminating free radicals from inside the cell [53].

In this study, treatment of AD rats with *B. carteri* daily for 8 weeks leads to significant increase in cortex and hippocampus homogenates of GSH, GSSG and SOD and significant decrease in LPO levels. The antioxidant activity of *B. serrata* (45 or 90 mg/kg b.w.) in the cerebrovascular system has been revealed [54]. *B. serrata* has potent antioxidant actions as shown by previous studies [55]. Forouzanfar *et al.* [56] reported that after cerebral ischemia, *B. serrata* extracts intake decrease MDA, and also increase GSH and SOD in cerebral cortex that suggests its antioxidant properties. Previous studies reported that boswellic acid has strong antioxidant activity [57]. The possibility of boswellic acid being an antioxidant and memory enhancer makes this agent an ideal neuroprotective agent. This finding is in agreement with others that high antioxidant agents increase learning and memory capacities of rats [58].

Treatment of AD rats with rivastigmine as a protective or therapeutic agent led to an improvement in the oxidative stress status, as represented in this study by a significant increase in the cortex and hippocampus homogenates of GSH, GSSG and SOD and significant decrease in LPO levels. Rivastigmine act via glutamergic mechanism, decreasing oxidative stress and restoring antioxidant defense [59].

Neuropathological examination of brain tissue sections of AD- rats showed neuronal degeneration and edema. Abnormal morphology in CA1 and DG of hippocampus characterized by degenerated shrunken cells with darkly stained nuclei. However, Al leads to hippocampus morphologically changes due to damage of neurons and gliosis, and this may affect various enzymes responsible for synthesis and destruction of the neurotransmitters [60]. Moreover, Al can pass the blood brain barrier, accumulate in brain and hippocampus, and produce neurotoxin damage [19].

In AD rats treated with *B. carteri*, or rivastigmine, hippocampus sections in CA1 and DG showed reduced degenerated cells. In addition, the hippocampus cells in CA1 and DG showed slight changes versus cells structure of rats treated with $AlCl_3$ alone. The hippocampus sections of CA1 in control, *B. carteri* extract and *B. carteri* + $AlCl_3$ treated rats revealed normal pyramidal cells with vesicular nuclei. Furthermore, the hippocampus sections of DG in control, *B. carteri* extract and *B. carteri*



+ AlCl_3 treated rats revealed normal granular cells with vesicular nuclei. Also, there was an increase in the thickness of the granular layer in *B. carteri* extract and (*B. carteri* + AlCl_3) treated groups.

Long-term oral intake of *B. serrata* in aged Wistar rat (100 mg/ kg/ day for 8 weeks) showed neuroprotective activity and increases CA1 pyramidal cells of hippocampus dendritic arbors [61]. Hosseini-Sharifabad *et al.* [62] reported that chronic administration with *B. serrata* (100 mg/ kg / day for 8 weeks) enhanced dendritic complexity in DG cells compared with control rats. Hippocampal granule cells of *B. serrata*- treated aged rats showed larger arbors, increased numerical branching density, more dendritic segments, and more dendritic spines versus controls. Karima *et al.* [24] revealed that boswellic acids increase neurite growth, branching and tubulin polymerization dynamics. Increase effect of boswellic acid on microtubule polymerization kinetics may be responsible for increasing axonal growth and branching. It prevents destruction of integrity of microtubule protein [63] due to neurodegeneration caused by aging [64]. *Boswellia* resin has many active substances in addition to boswellic acids. Incensole acetate, another substance of *Boswellia* resin also decrease hippocampal neuro-degeneration via its anti-inflammatory actions on brain [65]. Meanwhile, further studies need to know the biological mechanism by which *Boswellia*-induced neuritic enhancement.

In consistence with our histopathological results in *B. carteri* + AlCl_3 treated group, therapeutic and protective actions of *B. serrata* on AlCl_3 -induced AD in rats, have been found [11]. Chronic intake of methanolic extract of *B. serrate* had anti-inflammatory effects against neuro-inflammation manifested in AlCl_3 (AD rats model) [66]. It is probable that boswellic acids of frankincense might prevent starting of neuro-inflammation. Antioxidants prevent oxidative stress as they remove metal ions involved in neuronal plaque formation. Furthermore, in post-oxidative stress scenarios, antioxidant therapy hunts down free radicals and ROS and thus prevent neuronal degeneration [53].

Photomicrograph of brain of AD rats treated with rivastigmine revealed no histopathological alterations in the hippocampus. Coleman *et al.* [67] showed that rivastigmine treatment in a primary cell culture model preserve neuron, and preserve neuronal morphology and synaptic markers that are essential for normal neuronal action. Moreover, Bihaqi *et al.* [68] demonstrated normal histological appearance of the brain cells treated with rivastigmine tartrate. These authors stated that rivastigmine could reverse the histopathological alterations of the brain caused by Al.

CONCLUSION

From the result of this study we can conclude that *B. carteri* has a neuroprotective effect on rats with AD, prevents LPO rising, increases GSH, GSSG and SOD levels and improve memory and learning in the AD rat model. They also increase neurotransmitter formation in the brain as DA, NE and GABA and decreased AchE enzyme activity that destruct Ach which is excitatory neurotransmitter in the brain. They could ameliorate the neurodegenerative characteristics of AD. These results represented satisfactory therapeutic approaches for intervention against the progressive neurological damage associated with AD, with special reference to oxidative insults. Further clinical trials on humans are required to determine the efficacy of *B. carteri*, or one or more of its constituents, on neurodegenerative disorders.

REFERENCES

- [1] Fouladi, A. & Goli,S. Comparing working memory, verbal memory and keeping attention in the manic phase and depression in bipolar disorder. J Adv Pharm Edu Res 2018;8(2):82-84.
- [2] Hampel, H., Prvulovic, D., Teipel, S., Jessen, F., Luckhaus, C., Frölich, L., ...& Hoffmann, W. (2011). The future of Alzheimer's disease: the next 10 years. Progress in neurobiology, 95(4), 718-728.
- [3] Rashwan1, E.H. Kamel,M.M. El-lethey,H.S. Ciobica,A. El Iraqi,K.G.& Ahmed-Farid.O.A. Caffeine Ameliorating Effect on Anxiety and Depression in an Aluminum Chloride-induced Alzheimer's Disease Rat Model. International Journal of Pharmaceutical Research & Allied Sciences, 2018, 7(3):49-55.
- [4] Ahmed, M.Q. Alenazi, F.S. Fazaludeen,M.F. Shahid, S.M.& Kausar,M.A. Pathology and Management of Alzheimer's disease: A review. International Journal of Pharmaceutical Research & Allied Sciences, 2018, 7(2):30-42.
- [5] Kandimalla, R., Thirumala, V., & Reddy, P. H. (2017). Is Alzheimer's disease a type 3 diabetes? A critical appraisal. Biochimica et Biophysica Acta (BBA)-Molecular Basis of Disease, 1863(5), 1078-1089.
- [6] Prakash, A., Kalra, J., Mani, V., Ramasamy, K., & Majeed, A. B. A. (2015). Pharmacological approaches for Alzheimer's disease: neurotransmitter as drug targets. Expert review of neurotherapeutics, 15(1), 53-71.
- [7] Holtzman, D. M., Morris, J. C., & Goate, A. M. (2011). Alzheimer's disease: the challenge of the second century. Science translational medicine, 3(77), 77sr1-77sr1.

- [8] Siddiqui, M. Z. (2011). *Boswellia serrata*, a potential antiinflammatory agent: an overview. *Indian journal of pharmaceutical sciences*, 73(3), 255.
- [9] Rijkers, T., Ogbazghi, W., Wessel, M., & Bongers, F. (2006). The effect of tapping for frankincense on sexual reproduction in *Boswellia papyrifera*. *Journal of Applied Ecology*, 43(6), 1188-1195.
- [10] Hosseini Sharifabad, M., & Esfandiary, E. (2007). A morphometric study on CA3 hippocampal field in young rats following maternal administration of *Boswellia serrata* resin during gestation. *Iranian Journal of Basic Medical Sciences*, 10(3), 176-182.
- [11] Mahmoudi, A., Hosseini-Sharifabad, A., Monsef-Esfahani, H. R., Yazdinejad, A. R., Khanavi, M., Roghani, A., ...& Sharifzadeh, M. (2011). Evaluation of systemic administration of *Boswellia papyrifera* extracts on spatial memory retention in male rats. *Journal of natural medicines*, 65(3-4), 519.
- [12] Hosseini, M., Hadjzadeh, M. A. R., Derakhshan, M., Havakhah, S., Rassouli, F. B., Rakhshandeh, H., & Saffarzadeh, F. (2010). The beneficial effects of olibanum on memory deficit induced by hypothyroidism in adult rats tested in Morris water maze. *Archives of pharmacal research*, 33(3), 463-468.
- [13] Jalili, C., Salahshoor, M. R., Pourmotabbed, A., Moradi, S., Roshankhah, S. H., Darehdori, A. S., & Motaghi, M. (2014). The effects of aqueous extract of *Boswellia Serrata* on hippocampal region CA1 and learning deficit in kindled rats. *Research in pharmaceutical sciences*, 9(5), 351.
- [14] Yassin, N., El-Shenawy, S., Mahdy, K. A., Gouda, N., Marrie, A., Farrag, A., & Ibrahim, B. M. (2013). Effect of *Boswellia serrata* on Alzheimer's disease induced in rats. *J Arab Soc Med Res*, 8, 1-11.
- [15] Kivrak, E. G., Altunkaynak, B. Z., Alkan, I., Yurt, K. K., Kocaman, A., & Onger, M. E. (2017). Effects of 900-MHz radiation on the hippocampus and cerebellum of adult rats and attenuation of such effects by folic acid and *Boswellia sacra*. *Journal of Microscopy and Ultrastructure*, 5(4), 216-224.
- [16] Thenmozhi, A. J., Raja, T. R. W., Janakiraman, U., & Manivasagam, T. (2015). Neuroprotective effect of hesperidin on aluminium chloride induced Alzheimer's disease in Wistar rats. *Neurochemical research*, 40(4), 767-776.
- [17] Carageorgiou, H., Sideris, A. C., Messari, I., Liakou, C. I., & Tsakiris, S. (2008). The effects of rivastigmine plus selegiline on brain acetylcholinesterase, (Na⁺, K⁺)-, Mg²⁺-ATPase activities, antioxidant status, and learning performance of aged rats. *Neuropsychiatric disease and treatment*, 4(4), 687.
- [18] Staddon, J. E. (2016). *Adaptive behavior and learning*. Cambridge University Press.
- [19] Gol, M., Ghorbanian, D., Soltanpour, N., Faraji, J., & Pourghasem, M. (2017). Protective effect of raisin (currant) against spatial memory impairment and oxidative stress in Alzheimer disease model. *Nutritional neuroscience*, 1-9.
- [20] Kakad, V. D., Mohan, M., Kasture, V. S., & Kasture, S. B. (2008). Effect of *Vitis vinifera* on memory and behaviour mediated by monoamines. *Journal of Natural Remedies*, 8(2), 164-172.
- [21] Beheshti, S., & Aghaie, R. (2016). Therapeutic effect of frankincense in a rat model of Alzheimer's disease. *Avicenna journal of phytomedicine*, 6(4), 468.
- [22] Zaker, S. R., Beheshti, S., Aghaie, R., & Noorbakhshnia, M. (2015). Effect of olibanum on a rat model of Alzheimer's disease induced by intracerebroventricular injection of streptozotocin. *Physiology and Pharmacology*, 18(4), 477-489.
- [23] Beheshti, S., & Karimi, B. (2016). Frankincense improves memory retrieval in rats treated with Lipopolysaccharide. *Journal of HerbMed Pharmacology*, 5.
- [24] Karima, O., Riazi, G., Yousefi, R., & Movahedi, A. A. M. (2010). The enhancement effect of beta-boswellic acid on hippocampal neurites outgrowth and branching (an in vitro study). *Neurological Sciences*, 31(3), 315-320.
- [25] Onor, M. L., Trevisiol, M., & Aguglia, E. (2007). Rivastigmine in the treatment of Alzheimer's disease: an update. *Clinical interventions in aging*, 2(1), 17.
- [26] Gabrovska, K., Marinov, I., Godjevargova, T., Portaccio, M., Lepore, M., Grano, V., ...& Mita, D. G. (2008). The influence of the support nature on the kinetics parameters, inhibition constants and reactivation of immobilized acetylcholinesterase. *International journal of biological macromolecules*, 43(4), 339-345.
- [27] Zatta, P., Ibn-Lkhatyat-Idrissi, M., Zambenedetti, P., Kilyen, M., & Kiss, T. (2002). In vivo and in vitro effects of aluminum on the activity of mouse brain acetylcholinesterase. *Brain Research Bulletin*, 59(1), 41-45.
- [28] Struys-Ponsar, C., Guillard, O., & de Aguilar, P. V. D. B. (2000). Effects of aluminum exposure on glutamate metabolism: a possible explanation for

- its toxicity. *Experimental neurology*, 163(1), 157-164.
- [29] Kaizer, R. R., Corrêa, M. C., Spanevello, R. M., Morsch, V. M., Mazzanti, C. M., Gonçalves, J. F., & Schetinger, M. R. (2005). Acetylcholinesterase activation and enhanced lipid peroxidation after long-term exposure to low levels of aluminum on different mouse brain regions. *Journal of inorganic biochemistry*, 99(9), 1865-1870.
- [30] Zhang, J., Yang, J. Q., He, B. C., Zhou, Q. X., Yu, H. R., Tang, Y., & Liu, B. Z. (2009). Berberine and total base from rhizoma coptis chinensis attenuate brain injury in an aluminum-induced rat model of neurodegenerative disease. *Saudi medical journal*, 30(6), 760-766.
- [31] Kaizer, R. R., Correa, M. C., Gris, L. R. S., Da Rosa, C. S., Bohrer, D., Morsch, V. M., & Schetinger, M. R. C. (2008). Effect of long-term exposure to aluminum on the acetylcholinesterase activity in the central nervous system and erythrocytes. *Neurochemical research*, 33(11), 2294-2301.
- [32] Ibraheem, E. H. E., Moham, E. K. W., & Taher, M. A. (2011). Perturbation of brain neurotransmitters by aluminum in male rats and potential role of sage. *The Egyptian Journal of Experimental Biology (Zoology)*, 7(2), 249-259.
- [33] Delgado, M. R., Nystrom, L. E., Fissell, C., Noll, D. C., & Fiez, J. A. (2000). Tracking the hemodynamic responses to reward and punishment in the striatum. *Journal of neurophysiology*, 84(6), 3072-3077.
- [34] Martin, A. O., Mathieu, M. N., Chevillard, C., & Guérineau, N. C. (2001). Gap junctions mediate electrical signaling and ensuing cytosolic Ca²⁺ increases between chromaffin cells in adrenal slices: a role in catecholamine release. *Journal of Neuroscience*, 21(15), 5397-5405.
- [35] Burke, W. J., Li, S. W., Chung, H. D., Ruggiero, D. A., Kristal, B. S., Johnson, E. M., ... & Zahm, D. S. (2004). Neurotoxicity of MAO metabolites of catecholamine neurotransmitters: role in neurodegenerative diseases. *Neurotoxicology*, 25(1-2), 101-115.
- [36] Xu, Y., Stokes, A. H., Roskoski Jr, R., & Vrana, K. E. (1998). Dopamine, in the presence of tyrosinase, covalently modifies and inactivates tyrosine hydroxylase. *Journal of neuroscience research*, 54(5), 691-697.
- [37] Trombley, P. Q. (1998). Selective modulation of GABAA receptors by aluminum. *Journal of neurophysiology*, 80(2), 755-761.
- [38] Nayak, P., & Chatterjee, A. K. (2001). Differential responses of certain brain phosphoesterases to aluminium in dietary protein adequacy and inadequacy. *Food and chemical toxicology*, 39(6), 587-592.
- [39] Gonçalves, P. P., & Silva, V. S. (2007). Does neurotransmission impairment accompany aluminium neurotoxicity?. *Journal of Inorganic Biochemistry*, 101(9), 1291-1338.
- [40] Shuchang, H., Qiao, N., Piye, N., Mingwei, H., Xiaoshu, S., Feng, S., ... & Opler, M. (2008). Protective effects of gastrodia elata on aluminium-chloride-induced learning impairments and alterations of amino acid neurotransmitter release in adult rats. *Restorative neurology and neuroscience*, 26(6), 467-473.
- [41] Ota, M., & Houghton, P. (2005). Boswellic acid with acetylcholinesterase inhibitory properties from frankincense" 53rd annual congress organized by society of medicinal plants. *Societa Italiana di Fitochimica Florence*, 339.
- [42] Ota, M., & Houghton, P. J. (2008). Boswellic acids with acetylcholinesterase inhibitory properties from frankincense. *Natural Product Communications*, 3(1), 21-26.
- [43] Nakdook, W., Khongsombat, O., Taepavarapruk, P., Taepavarapruk, N., & Ingkaninan, K. (2010). The effects of Tabernaemontana divaricata root extract on amyloid β -peptide 25-35 peptides induced cognitive deficits in mice. *Journal of ethnopharmacology*, 130(1), 122-126.
- [44] Polinsky, R. J. (1998). Clinical pharmacology of rivastigmine: a new-generation acetylcholinesterase inhibitor for the treatment of Alzheimer's disease. *Clinical therapeutics*, 20(4), 634-647.
- [45] Andin, J., Enz, A., Gentsch, C., & Marcusson, J. (2005). Rivastigmine as a modulator of the neuronal glutamate transporter rEAAC1 mRNA expression. *Dementia and geriatric cognitive disorders*, 19(1), 18-23.
- [46] Yoneyama, M., Nishiyama, N., Shuto, M., Sugiyama, C., Kawada, K., Seko, K., ... & Ogita, K. (2008). In vivo depletion of endogenous glutathione facilitates trimethyltin-induced neuronal damage in the dentate gyrus of mice by enhancing oxidative stress. *Neurochemistry international*, 52(4-5), 761-769.
- [47] Deloncle, R., Huguet, F., Babin, P., Fernandez, B., Quellard, N., & Guillard, O. (1999). Chronic administration of aluminium L-glutamate in young mature rats: effects on iron levels and lipid

- peroxidation in selected brain areas. Toxicology letters, 104(1-2), 65-73.
- [48] Fraga, C. G., Oteiza, P. I., Golub, M. S., Gershwin, M. E., & Keen, C. L. (1990). Effects of aluminum on brain lipid peroxidation. Toxicology letters, 51(2), 213-219.
- [49] Shati, A. A., & Elsaid, F. G. (2009). Effects of water extracts of thyme (*Thymus vulgaris*) and ginger (*Zingiber officinale* Roscoe) on alcohol abuse. Food and chemical toxicology, 47(8), 1945-1949.
- [50] Mahdy, K., Shaker, O., Wafay, H., Nassar, Y., Hassan, H., & Hussein, A. (2012). Effect of some medicinal plant extracts on the oxidative stress status in Alzheimer's disease induced in rats. Eur Rev Med Pharmacol Sci, 16(3), 31-42.
- [51] Abd-Elhady, R. M., Elsheikh, A. M., & Khalifa, A. E. (2013). Anti-amnesic properties of Ginkgo biloba extract on impaired memory function induced by aluminum in rats. International Journal of Developmental Neuroscience, 31(7), 598-607.
- [52] Belaïd-Nouira, Y., Bakhta, H., Bouaziz, M., Flehi-Slim, I., Haouas, Z., & Cheikh, H. B. (2012). Study of lipid profile and parieto-temporal lipid peroxidation in AlCl₃ mediated neurotoxicity. Modulatory effect of fenugreek seeds. Lipids in health and disease, 11(1), 16.
- [53] Uttara, B., Singh, A. V., Zamboni, P., & Mahajan, R. T. (2009). Oxidative stress and neurodegenerative diseases: a review of upstream and downstream antioxidant therapeutic options. Current neuropharmacology, 7(1), 65-74.
- [54] Assimopoulou, A. N., Zlatanov, S. N., & Papageorgiou, V. P. (2005). Antioxidant activity of natural resins and bioactive triterpenes in oil substrates. Food chemistry, 92(4), 721-727.
- [55] Ammon, H. P. T. (2010). Modulation of the immune system by *Boswellia serrata* extracts and boswellic acids. Phytomedicine, 17(11), 862-867.
- [56] Forouzanfar, F., Hosseinzadeh, H., Ebrahimzadeh Bideskan, A., & Sadeghnia, H. R. (2016). Aqueous and ethanolic extracts of *Boswellia serrata* protect against focal cerebral ischemia and reperfusion injury in rats. Phytotherapy research, 30(12), 1954-1967.
- [57] Sabina, E. P., Indu, H., & Rasool, M. (2012). Efficacy of boswellic acid on lysosomal acid hydrolases, lipid peroxidation and anti-oxidant status in gouty arthritic mice. Asian Pacific journal of tropical biomedicine, 2(2), 128.
- [58] Ebrahimpour, S., Fazeli, M., Mehri, S., Taherianfard, M., & Hosseinzadeh, H. (2017). Boswellic Acid Improves Cognitive Function in a Rat Model Through Its Antioxidant Activity:- Neuroprotective effect of Boswellic acid. Journal of pharmacopuncture, 20(1), 10.
- [59] Shah, S., & Reichman, W. E. (2006). Treatment of Alzheimer's disease across the spectrum of severity. Clinical interventions in aging, 1(2), 131.
- [60] Lakshmi, B. V. S., Sudhakar, M., & Anisha, M. (2014). Neuroprotective role of hydroalcoholic extract of *Vitis vinifera* against aluminium-induced oxidative stress in rat brain. Neurotoxicology, 41, 73-79.
- [61] Hosseini-Sharifabad, M., & Esfandiari, E. (2015). Effect of *Boswellia serrata* gum resin on the morphology of hippocampal CA1 pyramidal cells in aged rat. Anatomical science international, 90(1), 47-53.
- [62] Hosseini-Sharifabad, M., Kamali-Ardakani, R., & Hosseini-Sharifabad, A. (2016). Beneficial effect of *Boswellia serrata* gum resin on spatial learning and the dendritic tree of dentate gyrus granule cells in aged rats. Avicenna journal of phytomedicine, 6(2), 189.
- [63] Karima, O., Riazi, G., Khodadadi, S., Yousefi, R., Mahnam, K., Mokhtari, F., ...& Moosavi-Movahedi, A. A. (2012). An in vitro study of the role of β -boswellic acid in the microtubule assembly dynamics. FEBS letters, 586(23), 4132-4138.
- [64] Himeda, T., Mizuno, K., Kato, H., & Araki, T. (2005). Effects of age on immunohistochemical changes in the mouse hippocampus. Mechanisms of ageing and development, 126(6-7), 673-677.
- [65] Moussaieff, A., Shein, N. A. A., Tsenter, J., Grigoriadis, S., Simeonidou, C., Alexandrovich, A. G., ...& Munoz, E. (2008). Incensole acetate: a novel neuroprotective agent isolated from *Boswellia carterii*. Journal of Cerebral Blood Flow & Metabolism, 28(7), 1341-1352.
- [66] Ahmed, H. H., Mohamed, E. M., & El-Dsoki, S. M. (2014). Evidences for the promising therapeutic potential of *Boswellia serrata* against Alzheimer's disease: pre-clinical study. Int J Pharm Pharm Sci, 6(11), 384-392.
- [67] Coleman, P., Federoff, H., & Kurlan, R. (2004). A focus on the synapse for neuroprotection in Alzheimer disease and other dementias. Neurology, 63(7), 1155-1162.
- [68] Bihagi, S. W., Sharma, M., Singh, A. P., & Tiwari, M. (2009). Neuroprotective role of *Convolvulus pluricaulis* on aluminium induced neurotoxicity in rat brain. Journal of ethnopharmacology, 124(3), 409-415.