



# Green Synthesis, Characterization and Antibacterial Activity of Zinc Nanoparticles Using Stem Extract of *Lawsonia inermis* Linn

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## ABSTRACT

In the production of nanoparticles, the bio-reduction properties of plant extracts are acknowledged worldwide to reduce the harmful effects of physical and chemical methods of synthesis of nanoscale metal particles. Zinc nanoparticles (ZnNPs) have been widely employed for various pharmacological applications. The present study recommends a green approach for the synthesis of zinc nanoparticles employing aqueous extract of stem of *Lawsonia inermis*. The biosynthesized ZnNPs were confirmed visually by the appearance of white color formation in the mixture. The prepared ZnNPs were characterized by UV-vis absorption, FTIR, SEM, XRD and AFM analysis. The UV-visible spectra showed that the absorption peak existed at 332nm. FTIR analysis revealed that phenolics, carboxylic acid, aromatic and amines present in the stem extract are responsible for the reduction and stabilization of the ZnNPs. SEM analysis demonstrated that the synthesized ZnNPs were spongy like shape. The XRD results indicated that the synthesized product showed hexagonal wurtzite phase and the average particle size is 17.80nm. The AFM image of ZnNPs exhibited a mixture of rod and spongy type of structure. The ZnNPs exhibited the antimicrobial activity against gram (+) bacteria, *Bacillus thuringiensis*, and *Streptococcus faecalis*. *L. inermis* may be used for the green synthesis of ultrafine nanoparticles of zinc for their antibacterial activities.

**Key Words:** Zinc nanoparticles, UV-Visible, FTIR, X-ray Diffraction, AFM.

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## INTRODUCTION

The growth of green processes for the blend of nanoparticles has been evolving into an essential branch of nanotechnology as green nanotechnology compacts with secure and eco-friendly ways for nanomaterials fabrication and this is considered as an alternative to conventional physical and chemical methods [1, 2]. For the past two decades, the research on synthesized nanomaterials and their distinct features is a new field of nanotechnology because of the wide applications in fields like chemistry, physics, biology, and medicine [3, 4]. Even- though

physical and chemical methods are more popular for nanoparticle synthesis, the use of toxic compounds, bounds their applications [5]. Definitely, over the past numerous years, plants, algae, fungi, bacteria, and viruses have been utilized for the production of metallic nanoparticles [6, 7]. Sometimes the amalgamation of nanoparticles using plants or parts of plants can establish advantages over other biological processes by eliminating the complicated processes of upholding microbial culture [8]. Using environmentally suitable plant extract and eco-friendly reducing and capping agents green nanoparticle synthesis has been reached. The use of plants for synthesis

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of nanoparticles is rapid, low cost, eco-friendly and a single-step method for the biosynthesis process [9]. Plant and plant materials have become a potential source for the synthesis of metallic nanoparticles recently. Many researchers have reported on the synthesis of metallic nanoparticles including silver, gold, titanium dioxide, tungsten oxide and copper oxide [10-14] using different plant materials. But reports on an amalgamation of zinc nanoparticles using plant materials are limited [15].

*Lawsonia inermis* Linn. (Family: Lythraceae) is a branched glabrous shrub or tiny tree (2-6 m in height), grown for its leaves while stem bark, roots, flowers, and seeds have also been employed in conservative medicine. This plant is a wide-reaching known superficial agent used to stain hair, skin, and nails [16]. The plant has reported the following properties: Analgesic, anti-inflammatory, antibacterial, wound healing, antimicrobial, hypoglycemic, immunostimulant, hepatoprotective, antifungal, antiviral, antiparasitic, antitrypanosomal, antioxidant, tuberculostatic, antidermatophytic, antifertility and anticancer properties.

The leaves of *Lawsonia inermis* are used to take care of poliomyelitis, measles among the Yoruba ethnic group of South Western Nigeria [17]. The seeds of henna have been accounted to possess deodorant action which is used in most cases of gynecological disorders like menorrhagia, vaginal discharge and leucorrhoea [18]. Henna is extensively used in the cosmetic industry as a dyeing agent as well in India [19]. Reports demonstrate that methanolic root extracts of *Lawsonia* are used in Nigeria as an antimalarial drug [20] in addition to abortifacient purposes [21]. There is ginger oil that is mixed with the powdered roasted seed to build a paste that is used for the managing ringworm. To heal wounds and clean wounds aseptically, the decoction of the leaves is used. [22]. *L.inermis* is also made use of by some individuals as 'blood tonic', thus implying its multifaceted application [20]. Hence the present study aims to develop a novel approach for the green synthesis of zinc nanoparticles using herbal plant *Lawsonia inermis* stem extract.

## MATERIALS AND METHODS

### Collection of Plant Material

*Lawsonia inermis* Linn. was collected from Pullavali, Pazhayakayal. The collected samples were cut into tiny fragments and shade dried in anticipation of the fracture is uniform and smooth. The dried material was granulated or powdered by using a blender, and sieved to get uniform particles by using sieve No. 60. The finishing uniform powder was utilized for the extraction of energetic constituents of the plant material. The plant samples were

found with the help of local flora and approved by the Botanical Survey of India, Southern Circle, Coimbatore, Tamil Nadu, India.

### Preparation of Extracts for Phytochemical Screening Hot Maceration Method Using Soxhlet Apparatus

Newly collected *L. inermis* stem was dried in shade, and then coarsely powdered in a blender. The coarse powder (100 g) was extracted successively with petroleum ether, benzene, ethyl acetate, methanol, ethanol and water, every 250 ml in a Soxhlet apparatus for 24 hrs. Through Whatman No.41 filter paper all the extracts were filtered. The entire extracts (petroleum ether, benzene, ethyl acetate, methanol, ethanol and aqueous) were focused on qualitative tests for the identification of different phytochemical constituents according to the standard procedures [23-25].

### Green Synthesis of Nanoparticles

#### Preparation of Stem Extract (Reducing Agent)

To prepare the stem extract, 20 gm of *L. inermis* stem was taken. Using double distilled water, the stem was thoroughly washed and then cut into fine pieces. Fine pieces were boiled in 100 ml double distilled water for 20 minutes at 60°C in a glass beaker. Using Whatman No. 1, the extract was filtered after boiling.

#### Preparation of Precursors

Precursors for zinc nanoparticles ( $\text{ZnCl}_2$ ) were purchased from Hi-media chemicals, India and prepared freshly. Precursor for preparing zinc nanoparticle was 0.02 M of zinc chloride using double distilled water.

#### Synthesis of Zinc Nanoparticles

10 ml aqueous solution of stem extract was slowly added to 10 ml of 0.02 M zinc chloride solution under constant stirring. Afterward, the mixture was treated with 1 M sodium hydroxide drop by drop. Soon after this, the mixture was changed into pale white. This was then kept in a magnetic stirrer for two hours. The white precipitate was obtained and dried in a hot air oven overnight at 60°C. Complete conversion of  $\text{Zn}(\text{OH})_2$  into Zn nanoparticles took place during drying.

## CHARACTERIZATION OF THE SYNTHESIZED ZINC NANOPARTICLE

### UV – Vis Spectroscopy

Ultraviolet-visible spectroscopy (UV-Vis) submits to absorption spectroscopy in the UV-visible spectral area. The zinc nanoparticles were characterized in a Shimadzu V 650 UV- Vis spectrophotometer. The scanning range for the samples was 300-700 nm. The double distilled water is made use as a blank reference.

### Fourier Transform Infra-Red Spectroscopy (FTIR)

The nanoparticles were illustrated utilizing a Fourier Transform Infrared Spectrophotometer (FTIR Thermoscientific iS5). Two milligrams of the sample was mixed with 100 mg Potassium bromide (KBr). Compressed to prepare a salt disc roughly 3 mm in diameter and the disc were at once kept in the sample holder. FTIR spectra were recorded in the absorption range between 400 and 4000  $\text{cm}^{-1}$ .

### Scanning Electron Microscope (SEM) Analysis

SEM is a kind of electron microscope that images a sample by scanning it with an elevated energy beam of electrons in a raster scan models. On a carbon-coated copper grid, this film of the sample was prepared by just dropping a very small amount of the sample on the grid. To remove the extra solution, a blotting paper was used and then the films on the SEM grid were permitted to dry by keeping it below a mercury lamp for 5 min.

### X-Ray Diffraction (XRD) Analysis

The particle size and nature of the zinc nanoparticles were determined using XRD. This was done using the Shimadzu XRD – 6000/6100 model with 30 kV, 30 mA with Cuka radians at 2 $\theta$  angle. X-ray powder diffraction is a fast analytical technique. This is chiefly used for phase identification of a crystalline material and can supply information on unit cell dimensions. The analyzed material is thinly ground, and the normal bulk composition is established. The particle or grain size of the zinc nanoparticles was determined using Debye Sherrer's equation.

$$D = 0.94 \lambda / B \cos \theta$$

### AFM Analysis

The surface topology of the synthesized zinc nanoparticles were studied by 1 $\mu\text{m}$  x 1  $\mu\text{m}$  Atomic Force Microscopy (AFM Nanosurf 2) analysis, 0.01 g synthesized nanoparticles were mixed with 20 ml of acetone and sonicated for 5-10 minutes using ultrasonicator. The solution was poured on a clean glass slide and was allowed to dry until all the acetone gets evaporated. To study this glass slide, the Atomic Force Microscopy is used in a non-contact mode. To process the captured image, XEI software was used.

### Antibacterial Activity

The antibacterial activity of synthesized nanoparticles was determined using a disc diffusion method [26]. The test bacteria *Bacillus thuringiensis*, *Streptococcus faecalis*, *Salmonella paratyphi*, and *Escherichia coli* was obtained from the Research Laboratory, Department of Microbiology, Bharathidasan University, Tiruchirapalli,

Tamil Nadu. The overnight incubated bacterial culture was spread over the freshly prepared nutrient agar plates. The 6 mm sterile disc (Hi media) was kept at the center and different concentrations of synthesized nanoparticles (40  $\mu\text{g/mL}$ , 80  $\mu\text{g/mL}$  and 100  $\mu\text{g/mL}$ ) were poured on disc and placed on the plate. The tetracycline disc (reference or positive control),  $\text{ZnCl}_2$  solution without extracts and plant aqueous extract were also kept and then incubated at 37 $^\circ\text{C}$  for 24h and after incubation the zone of inhibition was measured.

## RESULTS AND DISCUSSION

### Preliminary Phytochemical Analysis

The results of preliminary phytochemical screening of various solvent extracts of *L. inermis* stem were presented in Table 1. The methanol ethanol and aqueous extracts of *L. inermis* stem showed the presence of alkaloid, catechin, flavonoid, phenol, saponin, steroid, tannin, glycoside and xanthoprotein. Different phytochemicals were detected from *L. inermis* stem extracts which makes the plant useful for the treatment of different ailments. It also has the potential of providing useful drugs for human use.

### Synthesis of ZnNPs by Green Synthesis Process

The reaction was visible as a color change from light yellow to white within 24 hrs of the addition of reaction mixture Figure 1 showed the difference in the colour of extract and synthesized ZnNPs.

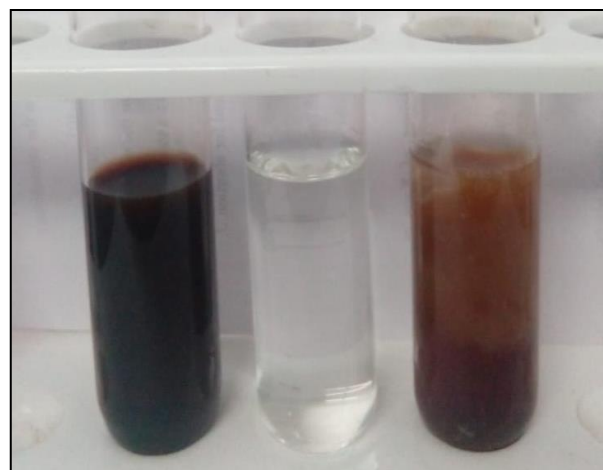


Figure 1: Synthesis of Zinc Nanoparticles from *L.inermis* Stem

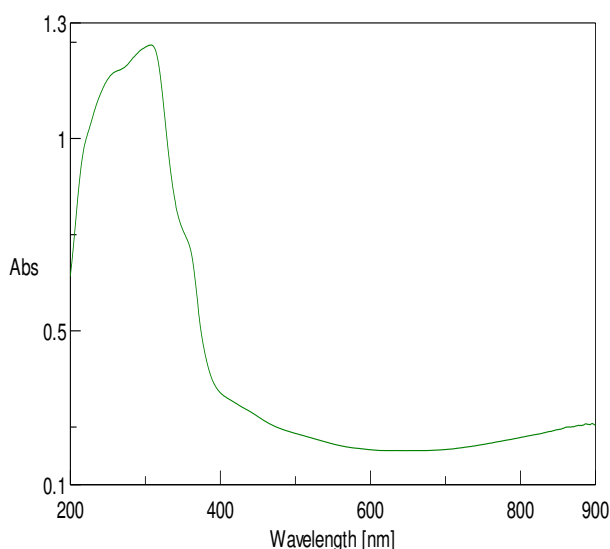
a) Plant Extract b) Zinc Chloride c) Zinc Nanoparticles

### Characterization of Synthesized ZnNPs

#### UV-Vis Spectroscopy Analysis

The nanoparticles were primarily characterized by UV-Vis spectrophotometer, which was proved to be a very useful technique for the analysis of nanoparticles. The absorption spectrum was recorded for the synthesis of ZnNPs in the range of 200-900nm by a UV-visible spectrophotometer.

The white color formation is the characteristic of ZnNPs formation and also depends on their Surface Plasmon Resonance (SPR). The spectrum showed the absorbance peak at 332nm corresponding to the characteristic of ZnNPs (Fig 2). Vimala *et al.* [27] reported on the absorbance spectra of ZnO nanoparticles which were recorded between 330nm and 370nm. The above findings were supported by present studies.

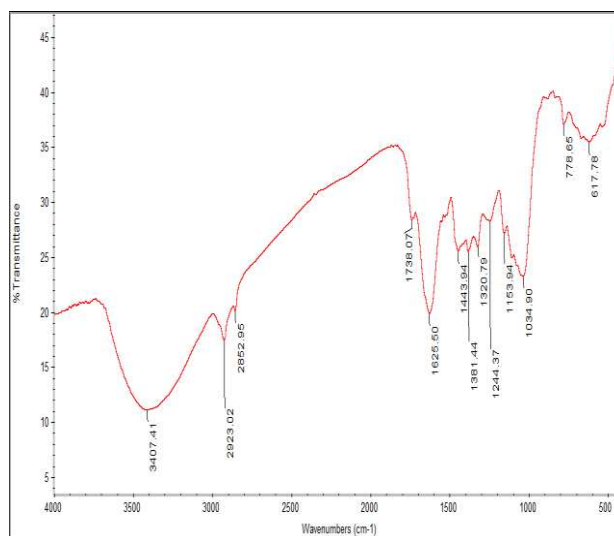


**Figure 2: UV-Visible Spectrum of Zinc Nanoparticles of *L. inermis* Stem**

#### Fourier Transform Infra-red Spectroscopy (FTIR)

The FTIR spectrum was utilized to recognize the functional groups of the lively components based on the peak value of the region of infrared radiation. The peak values of different compounds present with their frequencies and functional groups were shown in Table 2. The FTIR spectrum of the stem powder of *L. inermis*, (Figure 3) which clearly shows the peak at 3407  $\text{cm}^{-1}$  corresponds to the O-H stretching of a hydroxyl group, peak at 2923  $\text{cm}^{-1}$  represent O-H stretching of carboxylic acid, peak at 2852  $\text{cm}^{-1}$  assigned as C-H stretching of alkanes, peak at 1625  $\text{cm}^{-1}$  represent >N-H bond of a secondary amine, peak at 1443 and 1381  $\text{cm}^{-1}$  represent C-O stretching of esters and ethers, peak at 1224 and 1153  $\text{cm}^{-1}$  assigned as C-N stretching of aliphatic amines, peak at 778  $\text{cm}^{-1}$  represents C-H 'oop' of aromatics, peak at 617  $\text{cm}^{-1}$  assigned as C-Br stretching of alkyl halides. Figure 4 shows the FTIR spectrum of the biosynthesized zinc nanoparticles, peak at 3397  $\text{cm}^{-1}$  corresponds to the O-H stretching of alcoholic or phenolics, peak at 2923  $\text{cm}^{-1}$  represents O-H stretching of carboxylic acid, peak at 1597  $\text{cm}^{-1}$  assigned as C-C stretching of aromatics (in-ring), peak at 1493  $\text{cm}^{-1}$  represents C-C stretching of aromatics, peak at 1493  $\text{cm}^{-1}$  represent C-C stretching of aromatics, peak at 1383  $\text{cm}^{-1}$  corresponds to the C-H rock of alkanes,

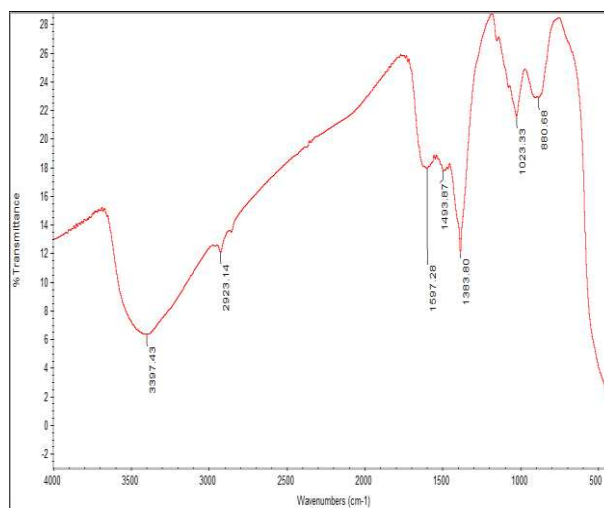
peak at 1023  $\text{cm}^{-1}$  represent C-N stretching of aliphatic amines, peak at 880  $\text{cm}^{-1}$  represent C-N 'oop' of aromatics.



**Figure 3: FT-IR Spectrum of Stem Powder of *L. inermis***

Figure 4 shows the FTIR spectrum of the biosynthesized zinc nanoparticles, peak at 3397  $\text{cm}^{-1}$  corresponds to the O-H stretching of alcoholic or phenolics, peak at 2923  $\text{cm}^{-1}$  represents O-H stretching of carboxylic acid, peak at 1597  $\text{cm}^{-1}$  assigned as C-C stretching of aromatics (in-ring), peak at 1493  $\text{cm}^{-1}$  represents C-C stretching of aromatics, peak at 1493  $\text{cm}^{-1}$  represent C-C stretching of aromatics, peak at 1383  $\text{cm}^{-1}$  corresponds to the C-H rock of alkanes, peak at 1023  $\text{cm}^{-1}$  represent C-N stretching of aliphatic amines, peak at 880  $\text{cm}^{-1}$  represent C-N 'oop' of aromatics.

FTIR measurement was performed to identify the biomolecules/organic molecules responsible for capping, reducing and stabilizing the ZnNPs present in the stem extract of *L. inermis*. The obtained results suggest the presence of various functional groups in the stem as well as in synthesized ZnNPs.



**Figure 4: FT-IR Spectrum of Zinc Nanoparticles of *L. inermis* Stem**



The stem extract has a variety of metabolites such as alcoholic or phenolics, carboxylic acid, alkane, secondary amine, aromatics, esters, ethers, aliphatic amines and alkyl halides which are responsible for the antioxidant or reducing property that aids in the immediate reduction of zinc ions into nano-structured ZnNPs. From the FTIR spectra of *L. inermis* stem extract and sample, there was an observed change in wavenumber of the functional groups that was due to the reduction and stabilization of metal group Zn [28].

### Scanning Electron Microscopy Analysis of ZnNPs (SEM)

The morphology and growth features of prepared ZnNPs were examined using scanning electron microscopy. Figure 5 showed the SEM image of the ZnNPs synthesized using *L. inermis* stem extract. The picture substantiates the spongy like the structure of the ZnNPs. From the SEM images, the crystallite size of ZnNPs synthesized using *L. inermis* stem extract was formed to be in the nanometre range.

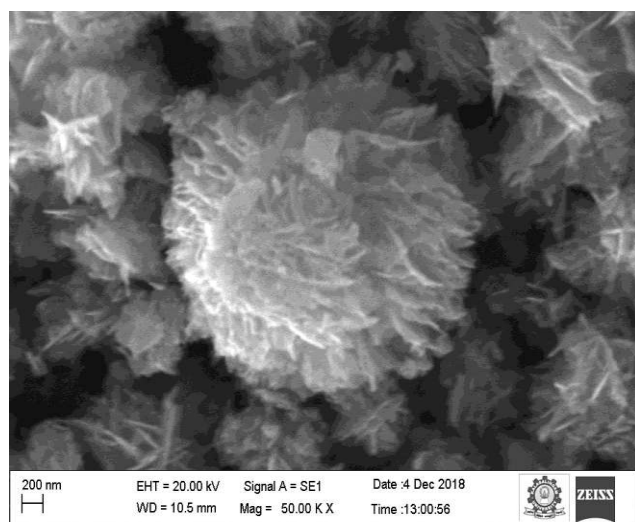


Figure 5: SEM Image of Zinc Nanoparticles of *L. inermis* Stem

### X-Ray Diffraction (XRD) Analysis of ZnNPs

XRD pattern of ZnNPs showed the peak corresponding to Bragg's diffraction signals from the crystal plants (111), (111), (111), (200), (210), (211), (211), (211) and (211). The intensity data were assembled starting from 20° to 80° over an array of 2θ. A definite line broadening of the XRD peaks indicates that the prepared materials consist of particles in the nanoscale range (Figure 6). The diffraction peaks located at 31.97°, 34.66°, 47.76°, 56.72°, 63.06°, 66.51°, 68.07°, 72.72° and 77.01° have been indexed as hexagonal wurtzite phase of ZnNPs. Furthermore, it proves that the synthesized nano-powder is pure as it does not have any characteristic XRD peaks other than ZnNPs. From the XRD peaks, the estimation of

average particle size was calculated using Scherrer's equation at 17.80nm.

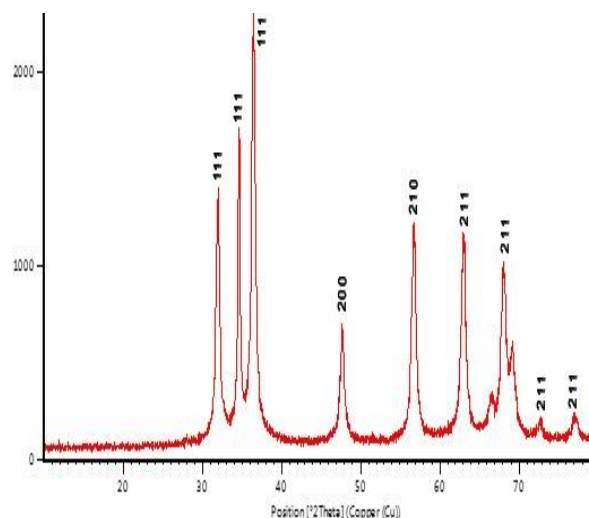


Figure 6: XRD Pattern of Synthesized Zinc Nanoparticles

### Atomic Force Microscopy (AFM) analysis of ZnNPs

The surface topology of the synthesized ZnNPs was studied by 1μm×1μm atomic force microscopy (AFM) analysis. AFM was used as the primary method to monitor ZnNPs dissolution and agglomeration pattern. The topography of micrographs clearly indicates that the formulated ZnNPs possess a mixture of rod and spongy shape as shown in Figure 7. The synthesized nanoparticles were stable in air and water and did not connect to any other associated compounds. Hence it exists highly dispersed nanoparticles.

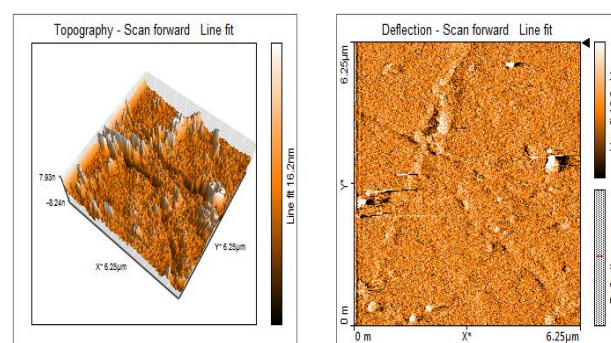


Figure 7: AFM Structure of Zinc Nanoparticles of *L. inermis* Stem

### Antibacterial Activity

Zinc nanoparticles (ZnNPs) were synthesized by using *L. inermis* stem extracts for the antibacterial activity against two gram (+) bacteria such as *Bacillus thuringiensis* and *Streptococcus faecalis* and two gram (-) bacteria like *Salmonella paratyphi* and *Escherichia coli*. ZnNPs have

been formed highly toxic against gram (+) bacteria such as *B. thuringiensis* and *Streptococcus faecalis* (Table 3).

What accounts for the enhanced antibacterial activity of nanoparticles is the increase in their surface area which is available for interaction. This enhances the bactericidal effect than the large-sized particles thereby imparting cytotoxicity to the microorganisms [29]. Researchers have not completely understood the mechanism that the nanoparticles used to penetrate the bacteria, but studies have shown that a change took place in its membrane morphology of the bacteria that were treated with zinc nanoparticles and this membrane morphology produced a noticeable increase in its permeability which affects proper transport through the plasma membrane [30, 31] leaving the bacterial cells incompetent of properly regulating transport through the plasma membrane, effecting into cell death. In the study, it was discovered that the zinc nanoparticles that have penetrated inside the bacteria caused damage by interacting with phosphorous and sulfur that contain compounds such as DNA [32, 33].

The result of numerous investigations have shown the possible mechanisms that involve the interaction of nanomaterials with the biological macromolecules. It is believed that microorganisms carry a negative charge while metal oxides carry a positive charge. This creates an 'electromagnetic' attraction between the microbe and the treated surface. ZnNPs synthesized from *L. inermis* stem extract proved to be a more potent antibacterial activity.

In conclusion, environmentally benign and low-cost synthesis of zinc nanoparticles can be achieved using the stem extract of *L. inermis*. The synthesized zinc nanoparticles exhibited good antibacterial activity. Therefore, nanoparticles of zinc in amalgamation with commercially available antibiotics could be utilized as an antibacterial agent after additional trials on experimental animals.

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**Table 1: Preliminary Phytochemical Screening of Different Solvent Extracts of Stem of *L. inermis***

Phytochemicals	Petroleum Ether	Benzene	Ethyl acetate	Methanol	Ethanol	Aqueous
Alkaloid	-	-	+	+	+	+
Anthraquinone	+	-	-	-	-	-
Catechin	-	+	-	+	-	+
Coumarin	-	+	-	-	+	-
Flavonoid	-	-	+	+	+	+
Phenol	+	-	+	+	+	+
Quinone	+	+	+	-	+	-
Saponin	-	-	+	+	+	+
Steroids	+	-	+	+	+	+
Tannin	-	+	-	+	+	+
Terpenoids	+	-	+	+	+	+
Sugar	+	-	+	-	+	-
Glycoside	+	-	+	-	-	-
Xanthoprotein	+	+	-	+	+	+
Fixed oil	+	+	-	-	+	-

+ Present - Absent

**Table 2: FT-IR Analysis of Powder and Synthesized Nanoparticles of *L. inermis***

S.No.	Frequency (cm <sup>-1</sup> )	Chemical Bond	Phytoconstituents Present	Peak Observed (Plant Powder)	Peak Observed (Zinc NPs)
1	3850-3500	O-H Stretch	Hydroxyl group	-	-
2	3500-3200	O-H Stretch	Alcohols or Phenols	3407	3397
3	3300-2500	O-H Stretch	Carboxylic acid	2923	2923
4	3000-2850	C-H Stretch	Alkanes	2852	-
5	2700-2250	NH <sup>+</sup> Stretch	Tertiary amine salt	-	-
6	1650-1550	>N-H bend	Secondary amine	1625	-
7	1600-1585	C-C Stretch(in ring)	Aromatics	-	1597
8	1500-1400	C-C Stretch	Aromatics	1443,1381	1493
9	1390-1350	C-H rock	Alkanes	-	1383
10	1360-1290	N-O Symmetric Stretch	Nitro Compound	-	-
11	1320-1000	C-O Stretch	Esters, Ethers	1320	-
12	1250-1020	C-N Stretch	Aliphatic amines	1153,1224	1023
13	910-665	N-H Wag	1°,2° amines	-	-
14	900-675	C-H"OOP"	Aromatics	778	880
15	690-400	C-Br Stretch	Alkyl halides	617	-

**Table 3: Antibacterial Activity of Synthesized Zinc Nanoparticles of *L.inermis***

Name of the Bacteria	Zone of Inhibition (mm)					
	<i>L. inermis</i> stem aqueous extract (100 µl)	Tetracycline (30 mcg/disc)	Zinc Chloride (100 µl)	ZnNPs Different Concentration		
				40µl	80 µl	100 µl
<i>Bacillus thuringiensis</i>	5	24	9	9	12	14
<i>Streptococcus faecalis</i>	4	27	10	10	11	12
<i>Salamonella paratyphi</i>	3	25	9	9	11	8
<i>Escherichia coli</i>	7	22	8	8	11	10