



Study of Oxidative Stress and Cytotoxicity in Reproductive Tissues of Adult Male Rats Exposed to Insecticides Proteus and Biscaya

Karamat Allah Solhjou^{1,2}, Seyed Ebrahim Hosseini^{2*}, Akbar Vahdati², Mohammad Amin Edalat Manesh²

¹ Department of Biology, Fars Science and Research Branch, Islamic Azad University, Fars, Iran.

² Department of Biology, Shiraz Branch, Islamic Azad University, Shiraz, Iran.

ABSTRACT

Insecticides, which are composed of endocrine disrupting compounds, lead to reduction of hormones and disturbance in the reproduction and growth of animals by interfering with endocrine systems. Also, reduced spermatogenesis activity and reproduction of animals are among other effects of insecticides. Considering that these compounds affect testicular activities and steroidogenesis by altering the activity of the relevant enzymes, and given the role of lactate dehydrogenase (LDH) enzyme in the function of spermatogenesis as well as presence of malondialdehyde (MDA) under oxidative stress conditions caused by tissue degradation, this study aimed to investigate changes in the two LDH and MDA enzymes influenced by the use of the insecticides Proteus and Biscaya, separately and in combination, and their effect on the sexual activity process in adult male rats.

Method: In this experimental study, 110 adult male 80-90-days-old Wistar rats with a mean weight of 200 g were selected and randomly divided into 11 equal groups including a control group (without receiving any substance), a control group (receiving distilled water), experimental groups 1, 2, 3 (respectively receiving 2.75, 5.5 and 11 mg/kg/Bw Proteus), experimental groups 4, 5, 6 (respectively receiving 1.5, 3, 6 mg/kg/Bw Biscaya), and experimental groups 7, 8, 9 (respectively receiving 1.5 mg/kg/Bw Biscaya+2.75 mg/kg/Bw Proteus, 3 mg/kg/Bw Biscaya+5.5 mg/kg/Bw Proteus, 6 mg/kg/Bw Biscaya+11 mg/kg/Bw Proteus). The LD50 method was used to evaluate the effect and amount of use of each of the insecticides. All the injections were performed intraperitoneally and the duration of the experiment was two weeks. At the end of the experiment period, blood samples were taken from the rats and the serum concentration of the LDH and MDA enzymes was measured using the ELISA kit. Statistical analysis was performed using SPSS version 16 and Duncan's test at a significant level of 0.05 ($P < 0.05$).

Findings: Comparison between the groups receiving different concentrations of the insecticides Proteus and Biscaya separately and in combination with the control group in the mean serum level of LDH and MDA showed that the maximum concentration of the insecticide Proteus and the medium and maximum concentrations of the insecticide Biscaya as well as different concentrations of a combination of these two types of insecticide significantly increased the level of LDH and MDA in a dose-dependent form.

Conclusion: Increased serum levels of LDH and MDA in the groups that received different doses of these insecticides separately and in combination were probably because the insecticides caused cytotoxicity and production of reactive oxygen species (ROS) in the dose-dependent form, which resulted in tissue degradation and elevated LDH. Moreover, by causing oxidative stress that was associated with increased MDA, these insecticides led to the production of nitric oxide (NO) and hydrogen peroxide (H₂O₂); these molecules, in turn, caused Leydig cell atrophy by their signaling function.

Key Words: LDH, MDA, Insecticide, Proteus, Biscaya, Rat.

eIJPPR 2017; 7(1):6-11

Corresponding author: Seyed Ebrahim Hosseini

Address: Ph.D, Department of Biology, Shiraz Branch, Islamic Azad University, Shiraz, Iran, 5kilometers to Sadra Town, Islamic Azad University Branch, Shiraz, Iran. Postcode: 71987-74731.

Phone : +089171183917

e-mail ✉ ebrahim.hosseini@yahoo.com

Relevant conflicts of interest/financial disclosures: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 25 September 2016; **Revised:** 15 February 2017; **Accepted:** 27 February 2017



INTRODUCTION

Endocrine disrupters are compounds that, in equilibrium, can cause changes in reproduction, growth or behavior, and thus affect animals or humans [1, 2]. These compounds destroy or inhibit endocrine systems, thus reducing or increasing the amount of hormones. Among endocrine disrupters are industrial and disposable products as well as pesticides including insecticides [3, 4]. Thiacloprid (THIA) from neonicotinoids and deltamethrin (DEL) from pyrethroids are among synthetic insecticides commonly used singly or in combination [5-9]. The insecticide Proteus, commercially known as DEL+THIA, and the insecticide Biscaya, commercially known as THIA, are two types of insecticide that have been widely used in agriculture in recent years; also, high levels of use of such pesticides have led to severe and varied effects on human health [10-13].

It has also been shown that these chemical compounds cause disturbances in the steroidogenesis mechanism in testicular tissues [14, 15]. It has been found that there is a high correlation between these factors (disrupting compounds) and reduction in sperm count, mobility and fertility as well as production of defective spermatozooids in animals [16]. Moreover, organochlorine pesticides have been reported to exert their effect by altering the activity of the enzymes involved in testicular activities [17, 18]. It has also been reported that the anti-androgenic effect caused by the use of glyphosate in animals increases the height of the epithelium of the tubes and reduces the cavity diameter of seminiferous tubules [19]. Another report states that imidacloprid increases intracellular space and atrophy of seminiferous tubules, reduces weight of the testicles, and reduces Leydig cells and their hypertrophy [20]. The use of dicofol at high and low doses has been reported to significantly reduce lactate dehydrogenase (LDH), which is a major enzyme for spermatogenesis, in testicular tissues. This reduction could be attributed to the interference of dicofol with energy metabolism in testicular tissues [21].

There are also reports on the impact of pyridabine on the increased levels of malondialdehyde (MDA) and nitric oxide (NO) [22]. Accordingly, it has been reported that testicular oxidative stress may be associated with decreasing testicular testosterone levels, and an increase in the NO level, similar to an oxidative index, results in inhibition of testosterone production [23].

On the other hand, it has been reported that pesticides lead to production of high levels of reactive oxygen species (ROS) in both extracellular and intracellular spaces, which is associated with a reduction in sperm count and fertility in humans and animals [24].

Therefore, considering the probable role of these toxins in the cytotoxicity of reproductive organs and their interference with the function of hormones [25], as well as the role of these pesticides in the production of high ROS levels in both extracellular and intracellular spaces followed by a reduction in sperm count [24] and

the richness of spermatozoa from unsaturated fatty acids that makes them suitable for oxidative stress [26], in this study, we attempted to measure the LDH level as a cytotoxicity marker and the MDA level as an oxidative stress indicator and also, to investigate the impact of use of the insecticides Proteus and Biscaya separately and in combination on these two enzymes.

MATERIALS AND METHOD

This is a completely randomized experimental study. The protocol of the study was established in accordance with the international regulations on the protection of laboratory animals and approved by the University's Ethics Committee under the number IR.miau13951206.

Animals and their grouping: In this research, all ethical considerations regarding the maintenance and use of laboratory animals were observed throughout the research period. In this experimental study, 110 adult male 80-90-days-old Wistar rats weighing 200-220 grams were used. The rats were kept at the animal breeding room of the Jahrom University of Medical Sciences for a period of one week to adapt to the environment. Throughout the study, the rats were maintained under a 12:12-h light:dark cycle at the environment temperature of 23±2°C and relative humidity of 50-55%. The rats were kept in shelves made of transparent makrolon polycarbonate and had free access to water and food.

The LD50 method was utilized to study the effect and amount of use of each of the insecticides [21]. The rats were randomly divided into 11 groups of 10, as described in Table 1.

Table 1. The different groups of the studied rats

Groups	Treatment
Control	Getting water and food naturally
Sham	Getting solvent of toxins (distilled water)
Experimental I	P 2/75 mg/kg/BW
Experimental II	P 5/5 mg/kg/BW
Experimental III	P 11 mg/kg/BW
Experimental IV	B 1/5 mg/kg/BW
Experimental V	B 3 mg/kg/BW
Experimental VI	B 6 mg/kg/BW
Experimental VII	B 1/5 + P 2/75 BW/kg/mg
Experimental VIII	B 3 + P 5/5 BW/kg/mg
Experimental IX	B 6 + P 11 BW/kg/mg

mg = milligram/ p = Proteus/ Kg = kilogram/ B = Biscaya/ BW = Body Weight

All the injections were performed intraperitoneally by insulin syringes. The injection rate was 0.4 cc in all the groups and the duration of the experiment was two weeks. The insecticides were manufactured by the Bayer CropScience Company.

Blood collection

One day after the last injection, blood samples were collected directly from the heart. Then, with the aid of a centrifuge machine (3000 rpm for 15 minutes), blood serums were collected and kept in the freezer at -20 °C until further use. To measure the LDH and MDA enzymes, the ELISA kit for rats manufactured by the Crystal Day Company of China (LOT = 2015 10-14-2016 10 13) was used.

Statistical analysis

A one-way analysis of variance (ANOVA) was used to analyze the data. According to the Kolmogorov-Smirnov test, the data distribution was normal and thus, analysis of parametric tests was used in the next stages. In cases where the statistical difference between the different groups was significant, Duncan's test was used to find out the difference between the means.

Statistical calculations were performed using SPSS version 16 and the significance level was considered to be $P < 0.05$. The data in the results section are given as Mean \pm SEM.

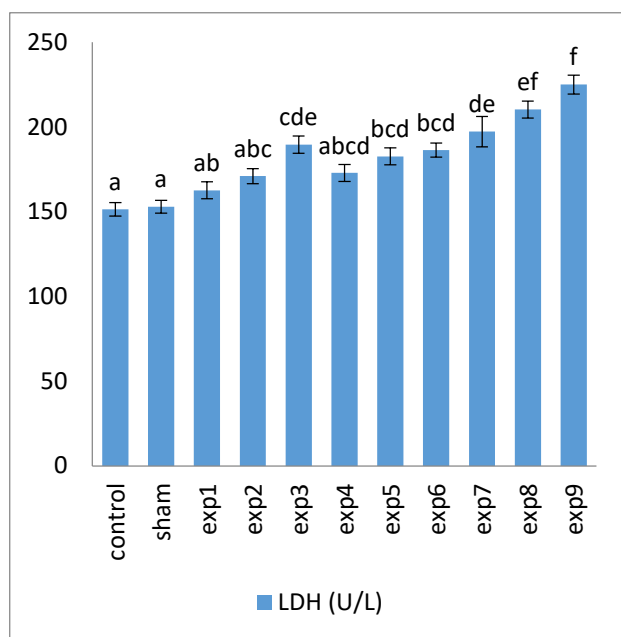


Figure 1. Evaluation of the LDH mean in the studied groups

- According to the Duncan's test, if there is at least one common letter in each column, groups in the columns are not significantly different from one another.
- U/l= Unit per Liter/ exp= experimental

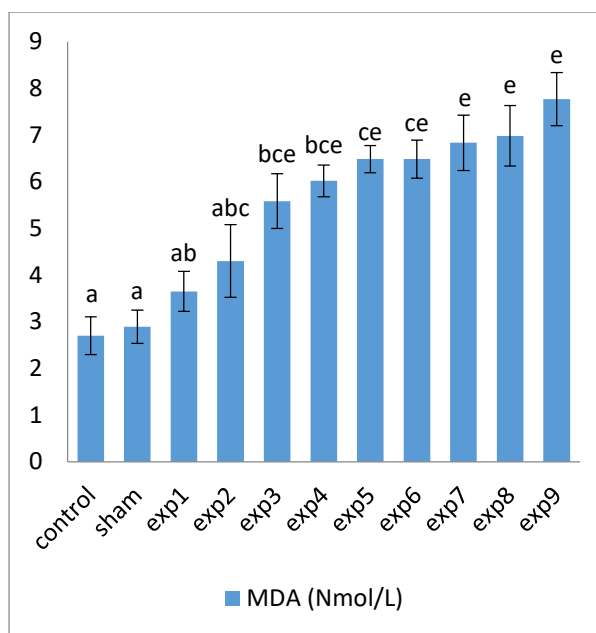


Figure 2. Evaluation of the MDA mean in the studied groups

- According to the Duncan's test, if there is at least one common letter in each column, groups in the columns are not significantly different from one another.
- Nmol/l= Nanomolar/ exp= experimental

Table 2. The parameters studied in the different groups (Mean \pm SEM)

Groups	MDA	LDH
Control	2.70 \pm .40 a	151.48 \pm 3.92 a
Sham	2.89 \pm .35 a	152.95 \pm 3.78 a
Experimental 1 (P 2.75 mg/kg)	3.65 \pm .42 ab	162.66 \pm 5.05 ab
Experimental 2 (P 5.5 mg/kg)	4.30 \pm .77 abc	171.00 \pm 4.38 abc
Experimental 3 (P 11 mg/kg)	5.58 \pm .58 bce	189.56 \pm 5.03 cde
Experimental 4 (B 1.5 mg/kg)	6.01 \pm .33 bce	172.90 \pm 4.95 abcd
Experimental 5 (B 3 mg/kg)	6.48 \pm .29 ce	182.66 \pm 4.95 bcd
Experimental 6 (B 6 mg/kg)	6.48 \pm .40 ce	186.36 \pm 4.15 bcd
Experimental 7 (P 2.75+B 1.5 mg/kg)	6.83 \pm .59 e	179.31 \pm 8.95 de
Experimental 8 (P 5.5+B 3 mg/kg)	6.98 \pm .65 e	210.30 \pm 5.03 ef
Experimental 9 (P 11+B 6 mg/kg)	7.76 \pm .56 e	225.05 \pm 5.52 f

- The mean of groups with dissimilar letters have significant difference.
- Data means are presented as Mean \pm SEM.
- The level of $p < 0.05$ is considered statistically significant.
- P = Proteus, B= Biscaya

FINDINGS

MDA: The comparison between the group receiving the maximum concentration of Proteus and the control group showed a significant increase in the mean serum concentration of MDA ($P < 0.05$). Also, the comparison of the groups receiving the minimum, medium and maximum concentrations of Biscaya with the control group revealed a significant increase in the mean serum concentration of MDA ($P < 0.05$).

In addition, the comparison between the groups receiving minimum, medium and maximum concentrations of a combination of Proteus and Biscaya with the control group showed a significant increase in the serum concentration of MDA, which was in a dose-dependent form so that the maximum concentration of the Proteus and Biscaya combination exerted a higher impact in the increase of the MDA serum concentration ($P < 0.05$).

LDH: A significant increase was observed in the mean serum concentration of LDH in the group receiving the maximum concentration of Proteus, as compared to the control group ($P < 0.05$). Also, the comparison between the groups that received different Proteus concentrations showed a significant increase in the LDH serum concentration in the groups receiving the minimum and medium concentrations of Proteus, as compared to the group receiving the maximum dose ($P < 0.05$).

Furthermore, the comparison between the groups receiving different Biscaya concentrations with the control group in terms of the LDH serum concentration showed that there was a significant difference between the groups receiving the medium and maximum Biscaya concentrations with the control group ($P < 0.05$). On the other hand, a significant increase was observed in the LDH level in a dose-dependent manner in the groups receiving the minimum, medium and maximum concentrations of a combination of Proteus and Biscaya, as compared to the control group ($P < 0.05$).

DISCUSSION

The results of this study showed that the maximum doses of Proteus alone and the medium and maximum doses of Biscaya alone, as well as different doses of a combination of Proteus and Biscaya significantly increased the mean serum levels of the MDA and LDH enzymes in the dose-dependent form. Increased serum levels of the MDA and LDH enzymes in this study are consistent with the results of other researchers. In the studies by Ebadi Manes, it has been reported that pyridabine led to the increased MDA and NO levels [22]. It has also been reported that presence of LDH is one way to detect cell death, and this cytoplasmic enzyme is released when the cell membrane is damaged, such as in cytotoxicity conditions [27]. Moreover, it has been reported that MDA is found in human and animal tissues as a lipid peroxidation end product, and is a

byproduct of biosynthesis of prostaglandins and thromboxanes [28].

Farag et al. reported that dimethoate has different effects on reproductive activities in male rats, including sperm density, mobility and vitality [29-31]. Also, Ngoula et al. reported that the pirimiphos-metyle insecticide reduced the number of Leydig cells [32].

Yet in other studies, it has been reported that testicular oxidative stress may be associated with lower testicular testosterone levels, and the increase in NO levels as an oxidative stress indicator results in inhibition of testosterone production [23].

CONCLUSION

Significant increase in the serum levels of the MDA and LDH enzymes between the groups that used different concentrations of Proteus and Biscaya separately or in combination with the control group was due to the degenerative effects of these pesticides [33], which caused atrophy and degeneration of the base cells as well as inhibition of spermatogenesis [29]. Hence, the increase in the LDH enzyme is probably due to cytotoxicity or cell death caused by these toxins. Moreover, these toxins may have caused Leydig cell apoptosis and atrophy in a dose-dependent manner [34]. The testicular oxidative stress associated with increasing NO levels has led to testosterone inhibition [23], and since NO and H₂O₂ can act as signaling molecules that results in oxidative cell death [35, 36], the LDH and MDA levels have increased. Therefore, considering that the reproductive hormones imbalance may contribute to reducing the amount of testicular antioxidants [37, 38], these toxins reduce testicular testosterone concentrations and, by producing high levels of ROS, stimulate lipid peroxidation and reduce enzymatic activities of antioxidants, and subsequently, cause apoptosis of base cells, degradation of spermatogenesis and elevation of LDH and MDA.

REFERENCES

- [1] Ormond G et al. Endocrine disruptors in the workplace, hair spray, folate supplementation, and risk of hypospadias: case-control study. *Environmental Health Perspectives* 2009; vol. 117, no. 2.
- [2] Lie D et al. Occupational exposure to bisphenol-A (BPA) and the risk of self-reported male sexual dysfunction. *Human Reproduction* 2010; vol. 25, no. 2.
- [3] Manif W et al. Effect of endocrine disruptor pesticides. *International Journal of Environmental Research and Public Health* 2011; vol. 8, no. 6.
- [4] Brophy J.T et al. Breast cancer risk in relation to occupations with exposure to carcinogens and endocrine disruptors: a Canadian case-control study. *Environmental Health* 2012; vol. 11: 87.
- [5] Zewen Liu, Xiangmel Yao and Yixi Zhang. Insect nicotinic acetylcholine receptors (nAChRs): important amino acid residue contributing to neonicotinoid



- insecticides selectivity and resistance. African Journal of Biotechnology 2008;7(25):4935-4939.
- [6] Masaru Shimomura, Maiko Yokota, Makoto Ihara, Miki Akamatus, David B. Sattelle, and Kazuhiko Matsuda. Role in the selectivity of neonicotinoids of Insect-specific Basic Residues in loop D of the Nicotinic Acetylcholine Receptor agonist binding site. Mol Pharmacol 2006;70:1255-1263.
- [7] Kazuhiko M, Masaru S, Makoto I, et al. Neonicotinoids show selective and diverse action on their nicotinic receptor target: electrophysiology, molecular biology, and receptor modeling studies. Biosci, Biotechnol, Biochem 2005;69(8):1442-1452.
- [8] Kazuhiko M, Satoshi K, Miki A, and David B.S. Diverse Actions and target-site selectivity of neonicotinoids: structural insights. The American Society for pharmacology and Experimental Therapeutics. Mol pharmacol 2009;76:1-10.
- [9] Sekeroglu V, Atl: Sekeroglu Z and kefelioglu H. Cytogenetic effects of commercial formulations of deltamethrin and/or thiacloprid on wistar rat bone marrow cells. Environmental Toxicology 2011; DoI: 10.1002/tox.20746.
- [10] Ghisari M, Bonefeld - Jorgensen EC. Impact of environmental chemicals on the thyroid hormone function in pituitary rat GH3 cells. Molecular and Cellular Endocrinology 2005;244:31-41.
- [11] Jesen AF, Peterson A and Granby K. Cumulative risk assessment of the intake of organophosphorus and carbamate pesticides in the Danish diet. Food Additives and Contaminants 2003;20:776-785.
- [12] Pitarch E, Serrano R, Lopez FJ and Hernandez F. Rapid multiresidue determination of organochlorine and organophosphorus compounds in human serum by solid phase extraction and gas chromatography coupled to tandem mass spectrometry. Analytical and Bioanalytical Chemistry 2003;376:189-179.
- [13] Sanghi R, Pillai MK, Jayalekshmi TR and Nair A. Organochlorine and Organophosphorus pesticide in breast milk from Bhopal, Madhya Pradesh, India. Human and Experimental Toxicology 2003;22:73-76.
- [14] Rhind SM. Endocrine disrupting compounds and farm animals their properties, actions and routes of exposure. Dom Anim Endocrinol 2002;23:179-178.
- [15] Mixed LG. Pesticides that confuse hormones. Pesticides Network UK. Briefing 2002;2:1-6.
- [16] Alm H, Tiemann U and Torner H. Influence of organochlorine pesticides on development of mouse embryos in vitro. Reprod Toxicol 1996;10:321-326.
- [17] Chitra KC, Latchoumycandane C and Mathur P.P. Chronic effect of endosulfan on the testicular functions of rat. Asian J. Andrology 1999;1:203-206.
- [18] Sinha N, Narayan R, Shancker R and Sexena D.K. Endosulfan-induced biochemical changes in the testis of rats. Vet. Hum. Toxicol 1995;37(6):547-549.
- [19] Romano R.M, Romano M.A, Bernardi M.M, Furtado P.V, Oliveira C.A. Pubertal exposure to commercial formulation of the herbicide glyphosate alters testosterone levels and testicular morphology. Arch Toxicol 2010;84:309-317.
- [20] Najafi G, Razi M, Hoshyar Shahmohammadloo A, Feyzi SS. The effect of chronic exposure with imidacloprid insecticide on fertility in mature male rats. International Journal of Fertility and Sterility 2010; 4(1):9-16.
- [21] Afaf A.El-K, Afrah F.S, Adel I.S, and Rania A.M. Chronic exposure of Dicofol promotes reproductive Toxicity in male rats. Life Science Journal 2010; Vol 7, NO 3.
- [22] Ebadi M G, Hasan Z S and Parivar K. The effects of pyridablin pesticide on the histomorphometric, hormonal alternations and reproductive functions of BALB/C mice. Iran J Basic Med Sci 2013;16(10):1055-64.
- [23] Mehta A, Sekhon C.P, Giri S, Orak J.K and Singh A.K. Attenuation of ischemia/reperfusion induced MAP kinase by N-acetyl cysteine, sodium nitroprusside and phosphoramidon. Molecular and Cellular Biochemistry 2002; vol.240. no.1-2, pp19-29.
- [24] Shorpe RM and Skakkebaek NE. Are oestrogen involved in falling sperm counts and disorders of the male reproductive tract? Lancet 1993;341:1392-1395.
- [25] Garcia AM., Occupational exposure and congenital malformations: a review of mechanism, methods, results. Amer. J. Indus. Med 1998;33:232-240.
- [26] Lenzi, A. Lipoperoxidation damage of spermatozoa polyunsaturated fatty acids (PUFA): Scavenger mechanisms and possible scavenger therapies. Front. Biosci 2000;5:1-15.
- [27] Francis Ka-Ming Chan, Kenta M, and Maria J D R. Detection Necrosis by Release of Lactate Dehydrogenase (LDH) Activity. Methods Mol Biol 2013;976:65-70. doi:10.1007/978-1-62703-290-7.
- [28] 039 IARC Monographs on the Evaluation of the Carcinogenic Risk of chemicals to Human IRAC 1985; vol.36. Allyl



- Compounds, Aldehydes, Exoxides, Lyon, pp. 163-177.
- [29] Farag AT, Ahmed F, Aswad E, Shaaban NA. Assessment of reproductive toxicity of orally administered technical dimethoate in male mice. *Reprod Toxicol* 2007; 23: 232-238.
- [30] Hwa-Young S, Yong-Bum K, Boo-Hyon K, Sung-Whan C, Chang-Su H, Jung-Koo R. Effects of 2-bromopropane on spermatogenesis in the sprague-dawley rat. *Reprod Toxicol* 1999; 13:179-187.
- [31] Afaf AM, Azza H. Evaluation of subchronic exposure of the male rat's reproductive system to the insecticide methomyl. *Saudi J Biol Sci* 2000; 7:2.
- [32] Ngoula F, Pierre W, Dongmo M, Kenfack A, Kamtchouing P, Tchoumboue. Effects of pirimiphos-methyl (an organophosphate insecticide) on the fertility of adult male rats. *Afr Health Sci* 2007; 7:3-9.
- [33] Sayým F. Histopathological effects of dimethoate on testes of rats. *Bull Environ Contam Toxicol* 2007; 78:479-484.
- [34] Jian Z, Lingling Z, Zheng L, Shuang W, and Liping X. Leptin level and oxidative stress contribute to obesity-Induced low Testicular Tissue. Handawi Publishing Corporation Oxidative Medicine and Cellular Longevity 2014; Vol Article ID 190945, 14 pages.
- [35] T. T. Turner and J. J. Lysiak, "Oxidative stress: a common factor in testicular dysfunction," *Journal of Andrology* 2008; vol. 29, no. 5, pp. 488-498.
- [36] Anand .H, Misro M.M, Sharma .S.B, and Prakash .S, "Cytoprotective effects of fruit pulp of *Eugenia jambolana* on H₂O₂-induced oxidative stress and apoptosis in rat Leydig cells invitro," *Andrologia* 2013; vol. 45, no. 3, pp. 145-157.
- [37] Zini .A and Schlegel P.N. "Effect of hormonal manipulation on mRNA expression of antioxidant enzymes in the rat testis," *Journal of Urology* 2003; vol. 169, no. 2, pp. 767-771.
- [38] Ghosh .D, Das U.B, Ghosh .S, Mallick .M, and Debnath .J, "Testicular gametogenic and steroidogenic activities in cyclophosphamide treated rat: a correlative study with testicular oxidative stress," *Drug and Chemical Toxicology* 2002; vol. 25, no. 3, pp. 281-292.